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## Nutraceutical Potential of Vitexin: A Flavone Glycoside

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### ABSTRACT

Vitexin is an apigenin flavone glycoside and it is extensively present in numerous edible and medicinal plants. It is considered as important as other flavonoids. It possessed a variety of biological properties, including anti-oxidation, anti-inflammation, anticancer, neuron-protective, cardio-protective, hepato-pancreatic protective effects. Other reported health relevant effects, includes fat reduction and glucose metabolism. The possible mechanism of protective effect through activation or inhibition of the different signaling pathways like AMPKa, Nrf-2, mTOR, PUMA, MMP, PARP, JNK, p38, Bcl-2/Bax ratio etc. The molecular targets of vitexin are very critical to utilize its potential effect as emerging chemopreventive and chemotherapeutic agent.

**Keywords:** Vitexin, ROS, Antioxidant, Anti-inflammatory, Anti-microbial, Anti-cancer.

### INTRODUCTION

Vitexin is an apigenin flavone glucoside (Apigenin-8-C-glucoside) has received great attention from scientists due to its diverse metabolic activities, like antioxidant, anti-microbial, anti-inflammatory, neuron-protective, anticancer and myocardial-protective effects [1-9]. Currently, it has been shown to exhibit several novel beneficial effects related to glucose homeostasis, fat metabolism, liver protective, antibacterial and antiviral [10-14]. Vitexins are present in various plants, including hawthorn [15], mung bean [16], beetroot [17], passion flowers [18, 19], otter flowers [20], bamboo [13], and gaillardia [21]. From ancient era, these plants have been utilized as food and medicine in Asia [22]. Still today it is widely utilized for the treatment of digestive related disorders, detoxification, dermal diseases, sun-stroke, inflammation and various other diseases and disorders [22]. Because of C-8 glucoside, vitexin causes a reduction of its bond dissociation enthalpy as compared to aglyconeapigenin, it has shown better free radical scavenger and antioxidant activity than apigenin. It has some derivatives too, such as isovitexin, rhamno-pyranosyl-vitexin, methyl-vitexin (isoembigenin), vitexin-2-O-rhamnoside, and vitexin-2-O-xyloside [17, 23]. The current literature, attempt has been made to discuss the nutraceutical potential of vitexin flavonoids.

### Nature source of vitexin

Scientifically, Vitexin known as apigenin-8-C-β-D-glycopyranoside or 5, 7, 4-trihydroxyflavone-8-glucoside. They are found in many plants, including *Vaccinium bracteatum* Thunb (Sea bilberry) [24], *Passiflora spp.* (Passion flower) [25, 26], *Pennisetum millet* (Candle millet) [27], *Vitex spp* (Chaste tree or Chaste berry) [28], *Bambusa vulgaris* (Bamboo leaves) [29], *Vignar adiata* (Mungbean) [30], *Triticum aestivum* (Wheat leaves) [31], *Mimosa pudica* (Mimosa) [32], *Crataegus spp* (Hawthorn) [33] among others.

### Delivery of vitexin

Vitexin can be consumed through edible plants, such as beetroot, or bamboo, mung beans and delivered to the target tissues at sufficiently high levels. Isolation of Vitexin can be from above mentioned plants as it is an integral part of that plants and transformed it into a bio-active constituent that can be incorporated into functional foods, supplements, or drugs. Molecular weight of Vitexin is about 432 Da, it's a small polar molecule. It can be delivered orally through foods, supplements, or pharmaceuticals in a number of ways. About solubility, it can be easily dissolved in aqueous-based foods, drinks and different pharma-based preparation. Again it could be converted into a powdered form that can then be incorporated into capsules or pills, tablets for supplement or pharmaceutical applications. The bioavailability and bioactivity of many plant based chemicals has been shown to be extended by controlling food matrix properties or by using colloidal delivery systems or [34-36].

## Pharmacokinetics of vitexin

Vitexin is not easily absorbed in the upper gastrointestinal tract. It has very low oral bioavailability due to early removal from blood. It is largely resistant to molecular transformations [37-39]. The metabolic transformation of this flavonoid glycoside occurs in the colon, where the micro-biota play principle roles in its degradation [40]. As the metabolic activity starts the Vitexin probably de-glycosylated initially and converted to 3-(4-hydroxyphenyl) propionic acid in the end. It is rapidly and widely distributed into various tissues but poor bioavailability. It is excreted mostly in the urine and bile [41, 42].

## Biological effects of vitexin

### Antioxidant effect

Oxidative stress causes excessive generation of reactive oxygen species (ROS), which causes damage to DNA, RNA and proteins of cells [43]. Chronic oxidative stress accelerates age-related dysfunctions, and play a major role in number of chronic disorders/diseases [9, 44, 45]. Vitexin has been reported as a potential antioxidant against free radicals [46]. It increases cell viability and reducing cellular injury by enhancing resistance against oxidative stress inducers. The capability to alleviate cellular reactive oxygen species (ROS) and malondialdehyde (MDA) levels has been revealed for vitexin [47-50]. This reduction in ROS and MDA by vitexin is attributed by increase of antioxidant enzyme activities, such as superoxide dismutase (SOD), heme-oxygenase, glutathione etc. Moreover the up-regulation of antioxidant response proteins like nuclear factor erythroid 2-related factor 2 or nuclear factor erythroid-derived 2-like 2 (NFE2L2) and 5'AMP-activated protein kinase/AMPK/5' adenosine monophosphate-activated protein kinase by vitexin play a major role against oxidative stress [47-50]. Out of above mentioned antioxidant proteins, nuclear factor erythroid 2 may regulate the expression of antioxidant proteins by which it protect against oxidative damage initiated by inflammation and cellular injury [51].

AMPK is an enzyme that plays a significant role in cellular energy homeostasis, largely to activate glucose and fatty acid uptake and oxidation. The main effect of AMPK activation is stimulation of liver and skeletal muscle fatty acid oxidation, keto-genesis, glucose uptake, inhibition of cholesterol bio-synthesis, triglyceride synthesis, adipocyte lipogenesis and modulation of insulin secretion by pancreatic B-cells [52]. In summary, vitexin can be considered as a potent antioxidant that prevent Reactive oxygen species induced degenerative damages such as hepatocytic injury, cardiac injury, neuro-degenerative disorder and insulin resistance both in vivo and in vitro [47-50].

### Anti-inflammatory effect

The anti-inflammatory properties of vitexin has gained major interest in recent research studies [13, 53-60]. Vitexin has been found to ameliorate lipopolysaccharide (LPS) induced inflammatory reactions [13, 55-61]. Actually, vitexin regulate the release of number of important inflammatory cytokines like tumor necrosis factor-Alpha (TNF- $\alpha$ ), Interleukin 1-b (IL-1b), and Interleukin-6 (IL-6) and enzymes (iNOS, MMP-1, MMP-3, and MMP-13) [56, 62-65]. Immuno-stimulating cytokines or bacterial pathogens activate inducible nitric oxide synthase (iNOS) and produced high concentrations of Nitric Oxide (NO) through the activation of inducible nuclear factors, including *nuclear factor kappa B* (NF- $\kappa$ B). Essentially, all members of the

matrix metallo-proteases (MMP) family have been linked to disease development, notably to tumour metastasis, chronic inflammatory reactions and the ensuing connective tissue damage as well as to nervous system disorders [66].

Immunoglobulin E (IgE) is important biomarker for allergic disorders [67]. Vitexin can inhibit the release of IgE into serum by inhibiting calcium release-activated calcium currents and b-hexosaminidase, which resulted in the amelioration of ovalbumin-induced allergic asthma [54]. In summary, vitexin can be considered to ameliorate organ injury, peritonitis, lung edema, inflammatory osteolysis etc [53, 56, 60, 68].

### Anti-cancer effect

Vitexin has also been shown to be a potent natural active ingredient in the inhibition of tumour cell genesis and tumor growth [69]. It exerts anti-neoplastic effects on cancer in various tissues, including the hepatic, intestinal, breast, dermal, lung, transisnal epithelial tissue (urinary) etc [69]. In recent years, there have been numerous studies mainly focusing on the inhibitory effects of vitexin on carcinogenesis by in-vitro route. There are different mechanism have been explored, it induced inhibition of cell proliferation by the prolongation of G2 cell-cycle arrest, cell apoptosis, the reduction of the Bcl-2/Bax ratio and the down-regulation of caspases [26, 70-75].

Cell susceptibility to apoptosis can be determined by Bcl-2/Bax ratio [76]. Bax/Bcl-2 ratio can affect tumor progression and aggressiveness through resistance of cancerous cells to apoptosis [76-78]. In other research it has been suggested that vitexin has MMP inhibitor property, which are important regulators of tumor proliferation and invasion [26] and up-regulate the poly (ADP-ribose) polymerase (PARP), an enzyme responsible for a various cellular processes such as damaged DNA repair, genomic stability and apoptosis (programmed cell death) [79].

In addition, vitexin up-regulated tumor suppressors: p53, PUMA (p53 up-regulated modulator of apoptosis) and several serine/threonine kinases [80, 81]. Regarding PUMA, it was identified as a transcriptional target of tumor suppressor (p53) and a potential apoptosis inducer in different neoplastic cells [82-84]. Again it is very interesting that PUMA found to be a critical mediator of p53-dependent and independent apoptosis induced by a wide variety of irritant like, genotoxic stress, deregulated tumour-oncogene expression, toxins, altered redox status, growth factor/cytokine withdrawal and infection [85].

In recent past, different active pharmaceuticals ingredient (API) targeting PUMA for cancer patients are emerging. These PUMA inducers target neoplastic cells, while PUMA inhibitors can be targeted to normal, healthy cells to reduce the unwanted effects of chemo and radiation therapy [85]. It has been also revealed that vitexin exerts it's anti-cancerous effects through inhibiting phosphatidyl-inositol-3-kinase (PI3K) [72, 75]. This pathway has been recently considered as an important mediator to cell proliferation, survival and oncogenesis [72, 75].

The mammalian target of rapamycin (mTOR, official known as the mechanistic target rapamycin) regulate various cellular functions. mTOR is basically a protein kinase encoded with MTOR gene. mTOR forms two different protein complex, known as mTORC-1 and mTORC -2. Out of these two forms, mTORC-1 generally regulates cell growth, cell mitotic activity, cell motility, protein synthesis, autophagy, and transcription [86, 87]. The activity of phosphoinositide 3-kinase (PI3K)/ mTOR pathway is dysregulated in

majority of neoplasm and has critical role in tumour development. Vitexin targeting the phosphoinositide 3-kinase (PI3K)/ mTOR signaling pathway so it would be an important therapeutic target agent in preventing carcinogenesis.

### Neuron-protective effect

The pathogenesis of neuronal disorders, including brain infarction, hypoxic-ischemic injury, epilepsy seizure, retinal damage, depression, slowness of movement (bradykinesia) and amnesia are closely related with the necrosis of neurons, reduced stress resistance and inflammatory condition in nervous tissues [47-49, 89-96]. Vitexin reduced aforesaid neuron disorders and improved the learning behaviors in laboratory animals mainly by decreasing Reactive Oxygen Substance (ROS) levels, decreasing the release of pro-inflammatory factors and promoting neuron survival [47-49, 89-96].

It also increased the neuron-cell viability by the up-regulation of the Bcl-2/Bax ratio and the down-regulation of cysteine-aspartic proteases *i.e* caspases [47, 90, 91, 94]. Considering the protein oxidation, lipid peroxidation and loss of mitochondrial membrane are typical neuronal cells damage inducer. Vitexin acted by reducing these oxidative stress substance level and thereby preventing injury to nervous [48, 49, 97]. In summary, vitexin has been shown to improve the neurogenesis and memory by alleviating the neuro-degenerative disorder, which lends support to the benefits of vitexin on sleep disorders [92].

### Cardio-protective effect

The pathogenesis of cardiovascular disorders, including infarction, atherosclerosis and myocardial cell necrosis are closely linked to inflammatory conditions, oxidative damage and endothelia cell injury [98-102]. Vitexin can protect the vascular endothelial cells from oxidized low density lipoproteins (ox-LDL) by enhancing the cell viability and physical stress resistance via the activation of AMPKa [50]. ox-LDL is the most damaging form of cholesterol that is generated when normal LDL cholesterol is modified by chemical interactions with free radicals particularly by myeloperoxidase (MPO) and reactive nitrogen species (RNS) [103,104].

The protective effect of vitexin in endothelia cells may also be attributed to the increased cell autophagy and up-regulation of relative genes, such as Beclin1 and light chain 3 II (LC3- II), as well as the down-regulation of p62 [50, 105]. Autophagy helped in tumor suppression by clearance of damaged cellular organelles and abnormal proteins. Basically it's a process to maintain cellular balance by the degradation of damaged organelles or misfolded proteins through the phagolysosome pathway. [101]

### Hepato-pancreatic protective effects

The pathogenesis of pancreatic gland injury and/or islet tissue damage usually causes insufficiency of beta cell secretion (insulin), which leads to impaired blood glucose absorption and finally increased glucose levels in blood [61,106]. Vitexin has demonstrated protective effect against pancreatic B- cell from toxicity induced by streptozotocin (STZ) or Lipo- polysaccharide (LPS) via the up-regulation of Nrf2 and antioxidants [107], as well as the inhibition of MAPKs (mitogen activated protein kinase or MAP kinase), including JNK (Jun N-terminal kinase) and p38 or p38 mitogen-activated protein kinases [10]. *Nrf2* induces cellular rescue pathways against oxidative injury, abnormal inflammatory and immune responses, as well as apoptosis [33], while MAPKs regulate cell functions like

cell proliferation, gene expression, differentiation, mitosis, cell survival, and apoptosis [108] and MAPKs including JNK and p38 are crucial regulators of cellular physiology, cell pathology, and many diseases including cancers [109].

Vitexin also increased glucose transporter type 4 (GLUT4) translocation from the cytoplasm to the cell membrane, suggesting the promotion of glucose uptake (absorption) [110]. GLUT4 is an insulin-regulated glucose transporter, responsible for insulin depended glucose uptake into fat and muscle cells [111]. It also improved diabetes-induced sexual dysfunction and fertility impairments in laboratory animals [112]. In this condition, vitexin therapy increased genital organs weight, reduced testicular pathological structure damage, restored spermatozoa quality, and improved sex hormonal balance [112]. In summary, vitexin have potential health benefits by protection against liver and pancreas injury.

### Metabolic (glucose and fat) effect

Vitexin have potential health benefits with respect to hepato-protection, glucose and fat metabolism [10-13, 96, 112]. As in the mechanism of glucose and fat metabolism, AMPKa was a key molecular target on vitexin-mediated fat reduction [11]. It activated AMPKa, a master in controlling fat accumulation [113], and inhibited CCAAT (cytosine-cytosine-adenosine-adenosine thymidine)/Enhancer Binding Protein Alpha (*C/EBP $\alpha$* ) and Fatty Acid Synthase (FAS), two contributors to lipogenesis and adipocyte differentiation [11]. Moreover, vitexin exerted protection against hepatic injury by down-regulating Alanine amino transferase, Asparted amino Transferase, Alkaline Phosphatase, and Lactate dehydrogenase enzymes involved in liver functions [12]. In summary, vitexin have potential health benefits by regulating the homeostasis of glucose and fat metabolism.

### Antimicrobial effects

Vitexin has shown inhibitory effects against gram negative bacteria, particularly *Pseudomonas aeruginosa* [68, 114]. *P. aeruginosa* can cause cystic fibrosis and infections in the urinary tract, kidney, and intestine through surface-attached biofilm on animal and human hosts [114]. Vitexin attenuate the formation of *P. aeruginosa* biofilms mainly through inhibiting cell adhesion ability, specifically by decreasing pathogen-swarming motility and down-regulating quorum-sensing regulator proteins [114].

### Antiviral effects

Vitexin also have ability to inhibit infection with influenza viruses [115, 116]. Many researches have shown that vitexin acted as an inhibitor for virus release and replication through inhibiting neuraminidase enzyme in influenza virus [115-117].

### CONCLUSION

Vitexin is obtained from the food sources. It is used as an active component with herbal supplement. The vitexin has multiple beneficial bioactivities. As a potent antioxidant, it prevent oxidative stress induced damages to the nervous, heart, liver-pancreas tissues and other systems with possible mechanism on molecular and cellular signaling. It produces different biological effects through activation or inhibition of the signaling pathways like AMPKa, Nrf-2, mTOR, PUMA, MMP, PARP, JNK, p38, Bcl-2/Bax ratio etc. It can enhance stress resistance, inflammatory responses, improvement of

energy homeostasis, inhibiting oncogenesis, detoxification etc. Conclusively, vitexin is a natural molecule with a variety of bioactivities which need further clinical trials to integrate it into effective functional food products.

#### Conflict of Interest

None declared.

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#### REFERENCES

1. Al-Jeboory A A, Dizaye KE H. Cardiovascular effects of vitexin. *Acta Pharmacol. Sin.* 2006;27:e146.
2. Choi HJ, Eun JS, Kim BG, Kim SY, Jeon H., Soh Y. Vitexin, a HIF-1 alpha a inhibitor, has anti-metastatic potential in PC12 cells. *Mol. Cells.* 2006;22(3):291-9.
3. Krcatovic E, Rusak G, Bezic N, Krajacic M. Inhibition of tobacco mosaic virus infection by quercetin and vitexin. *Acta Virol.* 2008;52:119-24.
4. An F, Yang, GD, Tian JM, Wang SH. Antioxidant effects of the orientin and vitexin in *Trollius chinensis* Bunge in Dgalactose-aged mice. *Neural Regen. Res.* 2012;7(33):2565-75.
5. Borghi SM, Carvalho TT, Staurengo-Ferrari L, Hohmann MSN, Pinge P, Casagrande R, Verri WA. Vitexin inhibits inflammatory pain in mice by targeting TRPV1, oxidative stress, and cytokines. *J. Nat. Prod.* 2013;76(6):1141-9.
6. Choi JS, Islam MN, Ali MY, Kim EJ, Kim YM, Jung HA. Effects of C-glycosylation on anti-diabetic, anti-alzheimer's disease and anti-inflammatory potential of apigenin. *Food Chem. Toxicol.* 2014;64:27-33.
7. Je HG, Hong SM, Je HD, Sohn UD, Choi YS, Seo SY, Min YS, Chung SJ, Shin YK, Lee TJ *et al.* The inhibitory effect of vitexin on the agonist-induced regulation of vascular contractility. *Pharmazie.* 2014;69(3):224-8.
8. Min JW, Hu JJ, He M, Sanchez RM, Huang WX, Liu YQ, Bsoul NB, Han S, Yin J, Liu WH *et al.* Vitexin reduces hypoxia-ischemia neonatal brain injury by the inhibition of HIF-1 alpha in a rat pup model. *Neuropharmacology.* 2015;99:38-50.
9. Xiao J, Capanoglu E, Jassbi AR, Miron A. Advance on the flavonoid C-glycosides and health benefits. *Crit. Rev. Food Sci. Nutr.* 2016;56(1):S29-S45.
10. Duncan SH, Schreiner M, Louis P, Maul R, Rohn S, Yan F, Yang Y, Yu L, Zheng X. Effects of C-glycosides from *Apios americana* leaves against oxidative stress during hyperglycemia through regulating mitogen-activated protein kinases and nuclear factor erythroid 2-related factor 2. *J. Agric. Food Chem.* 2017;65(34):7457-66.
11. Peng Y, Sun Q, Xu W, He Y, Jin W, Yuan L, Gao R. Vitexin ameliorates high fat diet-induced obesity in male C57BL/6J mice via the AMPKalpha-mediated pathway. *Food Funct.* 2019b;10(4):1940-7.
12. Wu H, Zhang G, Huang L, Pang H, Zhang N, Chen Y, Wang G. Hepatoprotective effect of polyphenol-enriched fraction from *Folium microcos* on oxidative stress and apoptosis in acetaminophen-induced liver injury in mice. *Oxid. Med. Cell. Longev.* 2017;2017:1-14.
13. Yang JH, Choi MH, Yang SH, Cho SS, Park SJ, Shin HJ, Ki SH. Potent anti-inflammatory and anti-adipogenic properties of Bamboo (*Sasa coreana* Nakai) leaves extract and its major constituent flavonoids. *J. Agric. Food Chem.* 2017;65(31):6665-73.
14. Fahmy NM, Al-Sayed E, Moghannem S, Azam F, El-Shazly M, Singab AN. Breaking down the barriers to a natural antiviral agent: Antiviral activity and molecular docking of *Erythrina speciosa* extract, fractions, and the major compound. *Chem. Biodivers.* 2020;17:e1900511.
15. Liang M, Xu W, Zhang W, Zhang C, Liu R, Shen Y, Li H, Wang X, Wang X, Pan Q *et al.* Quantitative LC/MS/MS method and in vivo pharmacokinetic studies of *Vitexin rhamnoside*, a bioactive constituent on cardiovascular system from hawthorn. *Biomed. Chromatogr.* 2007;21(4):422-9.
16. Luo J, Cai W, Wu T, Xu B. Phytochemical distribution in hull and cotyledon of adzuki bean (*Vigna angularis* L.) and mung bean (*Vigna radiate* L.), and their contribution to antioxidant, anti-inflammatory and anti-diabetic activities. *Food Chem.* 2016;201:350-60.
17. Ninfali P, Antonini E, Frati A, Scarpa ES. C-Glycosyl flavonoids from *Beta vulgaris* Cicla and betalains from *Beta vulgaris* rubra: Antioxidant, anticancer and anti-inflammatory activities-A review. *Phytother. Res.* 2017;31(6):871-84.
18. Colomeu TC, De Figueiredo D, Silva Matos P, Carvalho VD, Schumacher NSG, Abram DM, Fernandes LGR, Zollner RD. Effect of aqueous leaves extract from *Passiflora alata curtis* and vitexin, isoorientin in co-culture of min6/lymphocytes from nod mice in oxidative stress and cell death. *Ann. Nutr. Metab.* 2017;71:391-2.
19. Shuayprom A, Sanguansermisri D, Sanguansermisri P, Fraser IH, Wongkattiya N. Quantitative determination of vitexin in *Passiflora foetida* Linn. leaves using HPTLC. *Asian Pac. J. Trop. Biomed.* 2016;6(3):216-220.
20. Ngwoke KG, Ezenkwu C, Ajaghaku DL, Proksch P. Vitexin with its derivatives is responsible for the choline-mimetic properties of *Penianthus longifolius* extract which stimulates muscarinic receptors. *J. Nat. Prod.* 2017;7:231-6.
21. Moharram FA, El-Dib R, Marzouk MS, El-Shenawy SM, Ibrahim HA. New apigenin glycoside, polyphenolic constituents, anti-inflammatory and hepato-protective activities of *Gaillardia grandiflora* and *Gaillardia pulchella* aerial parts. *Pharmacogn. Mag.* 2017;13(50):S244-S249.
22. Ganesan K, Xu B J. A critical review on phytochemical profile and health promoting effects of mung bean (*Vigna radiata*). *Food Sci. Hum. Wellness.* 2018;7(1):11-33.
23. Praveena R, Sadasivam K, Kumaresan R, Deepha V, Sivakumar R. Experimental and DFT studies on the antioxidant activity of a C-glycoside from *Rhynchosia capitata*. *Spectrochim. Acta - A: Mol. Biomol. Spectrosc.* 2013;103:442-452.
24. Wang L, Zhang XT, Zhang HY, Yao HY, Zhang H. Effect of *Vaccinium bracteatum* Thunb. leaves extract on blood glucose and plasma lipid levels in streptozotocin-induced diabetic mice. *J. Ethnopharmacol.* 2010;130(3):465-469.
25. Zucolotto SM, Fagundes C, Reginatto FH, Ramos FA, Castellanos L, Duque C, Schenkel EP. Analysis of C-glycosyl flavonoids from South American *Passiflora* species by HPLC/DAD and HPLC-MS. *Phytochem. Anal.* 2012;23(3):232-239.

26. He M, Min JW, Kong WL, He XH, Li JX, Peng BW. A review on the pharmacological effects of vitexin and isovitexin. *Fitoterapia* 2016;115:74-85.
27. Gaitan E, Lindsay RH, Reichert RD, Ingbar SH, Cooksey RC, Legan J, Meydrech EF, Hill J, Kubota K. Antithyroid and goitrogenic effects of millet: role of C glycosyl flavones. *J. Clin. Endocrinol. Metab.* 1989;68(4):707-714.
28. Hajdu Z, Hohmann J, Forgo P, Martinek T, Dervarics M, Zupko I, Falkay G, Cossuta D, Mathe I. Diterpenoids and flavonoids from the fruits of *Vitex agnus-castus* and antioxidant activity of the fruit extracts and their constituents. *Phytother. Res.* 2007;21(4):391-394.
29. Lee HJ, Kim KA, Kang KD, Lee EH, Kim CY, Um BH, Jung SH. The compound isolated from the leaves of *Phyllostachys nigra* protects oxidative stress-induced retinal ganglion cells death. *Food Chem. Toxicol.* 2010;48(6):1721-1727.
30. Cao D, Li H, Yi J, Zhang J, Che H, Cao J, Yang L, Zhu C, Jiang W. Antioxidant properties of the mung bean flavonoids on alleviating heat stress. *PLoS One.* 2011;6(6):e21071.
31. Moheb A, Ibrahim RK, Roy R, Sarhan F. Changes in wheat leaf phenolome in response to cold acclimation. *Phytochemistry.* 2011;72(18):2294-2307.
32. Zhang J, Yuan K, Zhou WL, Zhou J, Yang P. Studies on the active components and antioxidant activities of the extracts of *Mimosa pudica* Linn. from southern China. *Pharmacogn. Mag.* 2011;7(25):35-39.
33. Ma LY, Liu RH, Xu XD, Yu MQ, Zhang Q, Liu HL. The pharmacokinetics of C-glycosyl flavones of Hawthorn leaf flavonoids in rat after single dose oral administration. *Phytomedicine.* 2010;17(8-9):640-645.
34. Goncalves RFS, Martins JT, Duarte CMM, Vicente AA, Pinheiro AC. Advances in nutraceutical delivery systems: From formulation design for bioavailability enhancement to efficacy and safety evaluation. *Trends Food Sci. Technol.* 2018;78:270-91.
35. McClements DJ, Xiao H. Designing food structure and composition to enhance nutraceutical bioactivity to support cancer inhibition. *Semin. Cancer Biol.* 2017;46:215-26.
36. Sayed N, Khurana A, Godugu C. Pharmaceutical perspective on the translational hurdles of phyto-constituents and strategies to overcome. *J. Drug Deliv. Sci. Technol.* 2019;53:101201.
37. Bai Y, Zhang Q, Wang B, Zhang M, Xu Y, Li S, Zhao Y, Yu Z. Plasma pharmacokinetics, bioavailability, and tissue distribution of four C-glycosyl flavones from mung bean (*Vigna radiate* L.) seed extracts in rat by ultrahigh-performance liquid chromatography- tandem mass spectrometry. *J. Agric. Food Chem.* 2017;65(27):5570-80.
38. Gao YC, Du Y, Ying ZM, Leng AJ, Zhang WJ, Meng YH, Li CY, Xu L, Ying XX, Kang TG. Hepatic, gastric and intestinal first-pass effects of vitexin-2-O-rhamnoside in rats by ultra-high-performance liquid chromatography. *Biomed. Chromatogr.* 2016;30(2):111-6.
39. Wang YJ, Han CH, Leng AJ, Zhang WJ, Xue HF, Chen YH, Yin JJ, Lu DR, Ying XX. Pharmacokinetics of vitexin in rats after intravenous and oral administration. *Afr. J. Pharmacy Pharmacol.* 2012;6(31):2368-2373.
40. Hein EM, Rose K, Van't Slot G, Friedrich AW, Humpf HU. Deconjugation and degradation of flavonol glycosides by pig cecal microbiota characterized by fluorescence in situ hybridization (FISH). *J. Agric. Food Chem.* 2008;56(6):2281-90.
41. Ninfali P, Angelino D. Nutritional and functional potential of *Beta vulgaris cicla* and *rubra*. *Fitoterapia.* 2013;89:188-199.
42. Xue HF, Ying ZM, Zhang WJ, Meng Y, Ying XX, Kang TG. Hepatic, gastric, and intestinal first-pass effects of vitexin in rats. *Pharm. Biol.* 2014;52(8):967-971.
43. Peng Y, Sun Q, Park Y. The bioactive effects of chicoric acid as a functional food ingredient. *J. Med. Food.* 2019c;22(7):645-52.
44. Papalia T, Barreca D, Panuccio MR. Assessment of antioxidant and cytoprotective potential of *Jatropha (Jatropha curcas)* grown in southern Italy. *Int. J. Mol. Sci.* 2017;18(3):e660.
45. Peng Y, Sun Q, Gao R, Park Y. AAK-2 and SKN-1 are involved in chicoric-acid-induced lifespan extension in *Caenorhabditis elegans*. *J. Agric. Food Chem.* 2019a;67(33):9178-86.
46. An F, Wang S, Yuan D, Gong Y, Wang S. Attenuation of oxidative stress of erythrocytes by the plant-derived flavonoids vitexin and apigenin. *Evid. Based Complement. Altern. Med.* 2016;2016:1-732.
47. Chen LL, Zhang B, Shan SQ, Zhao X. Neuroprotective effects of vitexin against isoflurane-induced neurotoxicity by targeting the TRPV1 and NR2B signaling pathways. *Mol. Med. Rep.* 2016;14(6):5607-13.
48. Malar DS, Prasanth MI, Shafreen RB, Balamurugan K, Devi KP. *Grewia tiliaefolia* and its active compound vitexin regulate the expression of glutamate transporters and protect Neuro-2a cells from glutamate toxicity. *Life Sci.* 2018a;203:233-41.
49. Malar DS, Suryanarayanan V, Prasanth MI, Singh SK, Balamurugan K, Devi KP. Vitexin inhibits A beta (25-35) induced toxicity in Neuro-2a cells by augmenting Nrf-2/HO-1 dependent antioxidant pathway and regulating lipid homeostasis by the activation of LXR-alpha. *Toxicol. In Vitro.* 2018b;50:160-71.
50. Zhang SL, Guo CL, Chen ZG, Zhang PY, Li JH, Li Y. Vitexin alleviates ox-LDL-mediated endothelial injury by inducing autophagy via AMPK signaling activation. *Mol. Immunol.* 2017;85:214-221.
51. Gold R, Kappos L, Arnold DL, Bar-Or A, Giovannoni G, Selmaj K *et al.* "Placebo-controlled phase 3 study of oral BG-12 for relapsing multiple sclerosis". *N. Engl. J. Med.* 2012;367(12):1098-107.
52. Winder WW, Hardie DG "AMP-activated protein kinase, a metabolic master switch: possible roles in type 2 diabetes". *Am. J. Physiol.* 1999;277(1): E1-10.
53. Jiang J, Jia Y, Lu X, Zhang T, Zhao K, Fu Z, Pang C, Qian Y. Vitexin suppresses RANKL-induced osteo-clastogenesis and prevents lipopolysaccharide (LPS)-induced osteolysis. *J. Cell. Physiol.* 2019;234(10):17549-12.
54. Kim YH, Oh T, Park E, Yim NH, Park K, Cho W, Ma J. Anti-inflammatory and anti-apoptotic effects of *Acer palmatum thumb.* extract, KIOM-2015EW, in a hyperosmolar-stress induced in vitro dry eye model. *Nutrients.* 2018a;10(3):e282.
55. Lee SJ, Hossaine MD, Park SC. A potential anti-inflammation activity and depigmentation effect of *Lespedeza bicolor* extract and its fractions. *Saudi J. Biol. Sci.* 2016;23(1):9-14.
56. Lu Y, Yu T, Liu JY, Gu LN. Vitexin attenuates lipopolysaccharide-induced acute lung injury by controlling the Nrf2 pathway. *PLoS One.* 2018;13(4):e0196405.
57. Rosa SIG, Rios-Santos F, Balogun SO, Martins DTD. Vitexin reduces neutrophil migration to inflammatory focus by down-regulating pro-inflammatory mediators via inhibition of p38, ERK1/2 and JNK pathway. *Phytomedicine.* 2016;23(1):9-17.
58. Schuster R, Holzer W, Doerfler H, Weckwerth W, Viernstein H, Okonogi S, Mueller M. *Cajanus cajan*- a source of PPARgamma activators leading to anti-inflammatory and cytotoxic effects. *Food Funct.* 2016;7(9):3798-3806.

59. Venturini CL, Macho A, Arunachalam K, de Almeida DAT, Rosa SIG, Pavan E, Balogun SO, Damazo AS, Martins DTD. Vitexin inhibits inflammation in murine ovalbumin induced allergic asthma. *Biomed. Pharmacother.* 2018;97:143-151.
60. Wang CL, Li TY, Zhang L, Zhu YT, Zhu KZ, Li ZY. Vitexin alleviates lipopolysaccharide-induced acute kidney injury via triggering AMPK/FOXO3a signaling pathway in newborn rats. *Lat. Am. J. Pharm.* 2019;38:558-564.
61. Wang F, Yin J, Ma Y, Jiang H, Li Y. Vitexin alleviates lipopolysaccharide-induced islet cell injury by inhibiting HMGB1 release. *Mol. Med. Rep.* 2017;15(3):1079-1086.
62. Kim HJ, Nam YR, Kim EJ, Nam JH, Kim WK. *Spirodela polyrhiza* and its chemical constituent vitexin exert antiallergic effect via ORAI1 channel inhibition. *Am. J. Chin. Med.* 2018b;46(06):1243-61.
63. Nikfarjam BA, Hajiali F, Adineh M, Nassiri-Asl M. Anti-inflammatory effects of quercetin and vitexin on activated human peripheral blood neutrophils-The effects of quercetin and vitexin on human neutrophils. *J. Pharmacopuncture.* 2017;20:127-31.
64. Xie CL, Li JL, Xue EX, Dou HC, Lin JT, Chen K, Wu HQ, Wu L, Xuan J, Huang QS. Vitexin alleviates ER-stress activated apoptosis and the related inflammation in chondrocytes and inhibits the degeneration of cartilage in rats. *Food Funct.* 2018;9(11):5740-5749.
65. Yang H, Huang J, Mao Y, Wang L, Li R, Ha C. Vitexin alleviates interleukin-1beta induced inflammatory responses in chondrocytes from osteoarthritis patients: Involvement of HIF-1alpha pathway. *Scand. J. Immunol.* 2019;90(2):e12773.
66. Klein T, Bischoff R. Physiology and pathophysiology of matrix metallo-proteases. *Amino Acids.* 2011;41(2):271-290.
67. Galli SJ, Tsai M. IgE and mast cells in allergic disease. *Nat. Med.* 2012;18(5):693-704.
68. Rath SN, Ray M, Pattnaik A, Pradhan SK. Drug target identification and elucidation of natural inhibitors for *Bordetella pertussis*: An in silico study. *Genomics Inform.* 2016;14(4):241-254.
69. Ganesan K, Xu BJ. Molecular targets of vitexin and isovitexin in cancer therapy: A critical review. *Ann. N. Y. Acad. Sci.* 2017;140(1):102-13.
70. Czemplik M, Mierziak J, Szopa J, Kulma A. Flavonoid C glucosides derived from flax straw extracts reduce human breast cancer cell growth in vitro and induce apoptosis. *Front. Pharmacol.* 2016;7:282-94.
71. Girish TK, Kumar KA, Prasada Rao UJS. C-glycosylated flavonoids from black gram husk: Protection against DNA and erythrocytes from oxidative damage and their cytotoxic effect on HeLa cells. *Toxicol. Rep.* 2016;3:652-63.
72. Liu XL, Jiang QF, Liu HM, Luo SX. Vitexin induces apoptosis through mitochondrial pathway and PI3K/Akt/mTOR signaling in human non-small cell lung cancer A549 cells. *Biol. Res.* 2019;52(1):e7.
73. Scarpa ES, Antonini E, Palma F, Mari M, Ninfali P. Antiproliferative activity of vitexin-2-O-xyloside and avenanthramides on CaCo-2 and HepG2 cancer cells occurs through apoptosis induction and reduction of pro-survival mechanisms. *Eur. J. Nutr.* 2018;57(4):1381-1395.
74. Scarpa ES, Emanuelli M, Frati A, Pozzi V, Antonini E, Diamantini G, Di Ruscio G, Sartini D, Armeni T, Palma F *et al.* Beta-cyanins enhance vitexin-2-O-xyloside mediated inhibition of proliferation of T24 bladder cancer cells dagger. *Food Funct.* 2016;7(12):4772-4780.
75. Zhang GN, Li DY, Chen H, Zhang JC, Jin XY. Vitexin induces G2/M-phase arrest and apoptosis via Akt/mTOR signaling pathway in human glioblastoma cells. *Mol. Med. Rep.* 2018;17:4599-4604.
76. Raisova M, Hossini AM, Eberle J, Riebeling C, Wieder T, Sturm I, Daniel PT, Orfanos CE, Geilen CC. The Bax/Bcl-2 ratio determines the susceptibility of human melanoma cells to CD95/Fas-mediated apoptosis. *J. Invest. Dermatol.* 2001; 117(2):333-40.
77. Wray CM, Ziogas A, Hinojosa MW, Le H, Stamos MJ, Zell JA. Tumor sub-site location within the colon is prognostic for survival after colon cancer diagnosis. *Dis. Colon. Rectum.* 2009;52(8):1359-66.
78. Hemminki K, Santi I, Weires M, Thomsen H, Sundquist J, Bermejo JL. Tumor location and patient characteristics of colon and rectal adenocarcinomas in relation to survival and TNM classes. *BMC Cancer.* 2010;10:688.
79. Herceg Z, Wang ZQ. "Functions of poly (ADP-ribose) polymerase (PARP) in DNA repair, genomic integrity and cell death". *Mutat. Res.* 2001;477(1-2):97-110.
80. Chen JF, Zhong JC, Liu YY, Huang Y, Luo F, Zhou YJ, Pan X, Cao SS, Zhang LL, Zhang YJ *et al.* Purified vitexin compound 1, a new neolignan isolated compound, promotes PUMA-dependent apoptosis in colorectal cancer. *Cancer Med.* 2018;7(12):6158-69.
81. Liu NA, Wang KS, Qi M, Zhou YJ, Zeng GY, Tao J, Zhou JD, Zhang J L, Chen X, Peng C. Vitexin compound1, a novel extraction from a Chinese herb, suppresses melanoma cell growth through DNA damage by increasing ROS levels. *J. Exp. Clin. Cancer Res.* 2018;37(1):e269.
82. Yu J, Zhang L, Hwang PM, Kinzler KW, Vogelstein B. PUMA induces the rapid apoptosis of colorectal cancer cells. *Mol. Cell.* 2001;7:673-82.
83. Nakano K, Vousden KH. PUMA, a novel pro-apoptotic gene, is induced by p53. *Mol. Cell.* 2001;7:683-94.
84. Han J, Flemington C, Houghton AB, Gu Z, Zambetti GP, Lutz RJ, Zhu L, Chittenden T. Expression of bbc3, a proapoptotic BH3-only gene, is regulated by diverse cell death and survival signals. *Proc. Natl. Acad. Sci. USA.* 2001;98(20):11318-23.
85. Yu J, Zhang L. PUMA, a potent killer with or without p53. *Oncogene.* 2008; 27(Spl 1):S71-83.
86. Jacinto E, Loewith R, Schmidt A, Lin S, Ruegg MA, Hall A, Hall MN. "Mammalian TOR complex 2 controls the actin cytoskeleton and is rapamycin insensitive". *Nat. Cell Biol.* 2004;6(11):1122-8.
87. Lipton JO, Sahin M. "The neurology of mTOR". *Neuron.* 2014; 84(2):275-291.
88. Porta C, Paglino C, Mosca A. Targeting PI3K/Akt/mTOR signaling in cancer. *Front. Oncol.* 2014;4:64.
89. Aseervatham, GS, Suryakala U, Doulethunisha SS, Bose PC, Sivasudha T. Expression pattern of NMDA receptors reveals antiepileptic potential of apigenin 8-C-glucoside and chlorogenic acid in pilocarpine induced epileptic mice. *Biomed. Pharmacother.* 2016;82:54-64.
90. Hu M, Li FM, Wang WD. Vitexin protects dopaminergic neurons in MPTP-induced Parkinson's disease through PI3K/Akt signaling pathway. *Drug Des. Devel. Ther.* 2018;12:565-73.
91. Jiang J, Dai JC, Cui H. Vitexin reverses the autophagy dysfunction to attenuate MCAO-induced cerebral ischemic stroke via mTOR/Ulk1 pathway. *Biomed. Pharmacother.* 2018;99:583-90.
92. Kim GH, Lim K, Yang HS, Lee JK, Kim Y, Park SK, Kim SH, Park S, Kim TH, Moon JS *et al.* Improvement in neurogenesis and memory function by administration of *Passiflora incarnata*

- L. extract applied to sleep disorder in rodent models. J. Chem. Neuroanat. 2019;98:27-40.
93. Luo WD, Min JW, Huang WX, Wang X, Peng YY, Han S, Yin J, Liu WH, He XH, Peng BW. Vitexin reduces epilepsy after hypoxic ischemia in the neonatal brain via inhibition of NKCC1. J. Neuroinflammation. 2018;15(1):e186.
94. Lyu ZP, Cao J, Wang J, Lian HM. Protective effect of vitexin reduces sevoflurane-induced neuronal apoptosis through HIF-1 alpha, VEGF and p38 MAPK signaling pathway in vitro and in newborn rats. Exp. Ther. Med. 2018;15:3117-23.
95. Min JW, Kong WL, Han S, Bsoul N, Liu WH, He XH, Sanchez RM, Peng BW. Vitexin protects against hypoxic ischemic injury via inhibiting Ca2p/Calmodulin-dependent protein kinase II and apoptosis signaling in the neonatal mouse brain. Oncotarget. 2017;8(15):25513-24.
96. Nurdiana S, Goh YM, Hafandi A, Dom SM, Nur Syimal'ain A, Noor Syaffinaz NM, Ebrahimi M. Improvement of spatial learning and memory, cortical gyrification patterns and brain oxidative stress markers in diabetic rats treated with *Ficus deltoidea* leaf extract and vitexin. J. Tradit. Complement. Med. 2018;8(1):190-202.
97. Cui YH, Zhang XQ, Wang ND, Zheng MD, Yan J. Vitexin protects against ischemia/reperfusion-induced brain endothelial permeability. Eur. J. Pharmacol. 2019;853:210-9.
98. Bai Y, Chang J, Xu Y, Cheng D, Liu H, Zhao Y, Yu Z. Antioxidant and myocardial preservation activities of natural phytochemicals from mung bean (*Vigna radiata* L.) seeds. J. Agric. Food Chem. 2016;64(22):4648-55.
99. Che X, Wang X, Zhang JY, Peng CF, Zhen YL, Shao X, Zhang GL, Dong LY. Vitexin exerts cardio-protective effect on chronic myocardial ischemia/reperfusion injury in rats via inhibiting myocardial apoptosis and lipid peroxidation. Am. J. Transl. Res. 2016;8(8):3319-28.
100. Cheng D, Wang R, Wang C, Hou L. Mung bean (*Phaseolus radiatus* L.) polyphenol extract attenuates aluminum induced cardiotoxicity through an ROS-triggered Ca2p/JNK/NFkappa B signaling pathway in rats. Food Funct. 2017;8(2):851-9.
101. Glick D, Barth S, Macleod KF. Autophagy: Cellular and molecular mechanisms. J. Pathol. 2010;221(1):3-12.
102. Sun Z, Yan B, Yu WY, Yao XP, Ma XJ, Sheng GL, Ma Q. Vitexin attenuates acute doxorubicin cardiotoxicity in rats via the suppression of oxidative stress, inflammation and apoptosis and the activation of FOXO3a. Exp. Ther. Med. 2016;12(3):1879-1884.
103. Gao S, Zhao D, Wang M, Zhao F, Han X, Qi Y, Liu J. Association Between Circulating Oxidized LDL and Atherosclerotic Cardiovascular Disease: A Meta-analysis of Observational Studies. Can. J. Cardiol. 2017;33(12):1624-1632.
104. Suci CF, Prete M, Ruscitti P, Favoino E, Giacomelli R, Perosa F. Oxidized low density lipoproteins: The bridge between atherosclerosis and autoimmunity. Possible implications in accelerated atherosclerosis and for immune intervention in autoimmune rheumatic disorders. Autoimmun. Rev. 2018;17(4):366-375.
105. Wirawan E, Lippens S, Vanden-Berghe T, Romagnoli A, Fimia GM, Piacentini M, Vandenabeele P. Beclin1: A role in membrane dynamics and beyond. Autophagy. 2012;8(1):6-17.
106. Peng Y, Sun Q, Park Y. Chicoric acid promotes glucose uptake and Akt phosphorylation via AMP-activated protein kinase a-dependent pathway. J. Funct. Foods. 2019d;59:8-15.
107. Nurdiana S, Goh YM, Ahmad H, Dom SM, Syimal'ain Azmi N, Noor Mohamad Zin, NS, Ebrahimi M. Changes in pancreatic histology, insulin secretion and oxidative status in diabetic rats following treatment with *Ficus deltoidea* and vitexin. BMC Complement Altern. Med. 2017;17(1):e290.
108. Pearson G, Robinson F, Beers Gibson T, Xu BE, Karandikar M, Berman K, Cobb MH "Mitogen-activated protein (MAP) kinase pathways: regulation and physiological functions". Endocr. Rev. 2001;22(2):153-83.
109. Mendelson KG, Contois LR, Tevosian SG, Davis RJ, Paulson KE. Independent regulation of JNKp38 mitogen-activated protein kinases by metabolic oxidative stress in the liver. Proc. Natl. Acad. Sci. USA. 1996;93:12908-12913.
110. Seyedan A, Mohamed Z, Alshagga MA, Koosha S, Alshawsh MA. *Cynometra cauliflora* Linn. attenuates metabolic abnormalities in high-fat diet-induced obese mice. J. Ethnopharmacol. 2019;236:173-182.
111. Stockli JS, Fazakerley DJ, James DE. GLUT4 exocytosis. J. Cell. Sci. 2011;124(24):4147-59.
112. Li ZM, Liu N, Jiang YP, Yang JM, Zheng J, Sun M, Li YX, Sun T, Wu J, Yu JQ. Vitexin alleviates streptozotocin induced sexual dysfunction and fertility impairments in male mice via modulating the hypothalamus-pituitary-gonadal axis. Chem. Biol. Interact. 2019;297:119-29.
113. Sun Q, Lin J, Peng Y, Gao R, Peng Y. Flubendiamide enhances adipogenesis and inhibits AMPKa in 3T3-L1 adipocytes. Molecules. 2018;23(11):e2950.
114. Das MC, Sandhu P, Gupta P, Rudrapaul P, De UC, Tribedi P, Akhter Y, Bhattacharjee S. Attenuation of *Pseudomonas aeruginosa* biofilm formation by vitexin: A combinatorial study with azithromycin and gentamicin. Sci. Rep. 2016;6(1):23347.
115. Ding F, Liu J. Qualitative and quantitative analysis for the chemical constituents of *Tetrastigma hemsleyanum* diels et gilg using ultra-high performance liquid chromatography/hybrid quadrupole orbitrap mass spectrometry and preliminary screening for anti-influenza virus components. Evid. Based Complement Alternat. Med. 2019;2019:9414926.
116. Sadati SM, Gheibi N, Ranjbar S, Hashemzadeh MS. Docking study of flavonoid derivatives as potent inhibitors of influenza H1N1 virus neuraminidase. Biomed. Rep. 2019;10(1):33-38.
117. Li YL, Ma SC, Yang YT, Ye SM, But PP. Antiviral activities of flavonoids and organic acid from *Trollius chinensis* Bunge. J. Ethnopharmacol. 2002;79(3):365-8.

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