

The Journal of Phytopharmacology

(Pharmacognosy and phytomedicine Research)



Research Article

ISSN 2320-480X
JPHYTO 2025; 14(3): 165-169
May- June
Received:23-05-2025
Accepted: 24-07-2025
Published: 31-07-2025
©2025, All rights reserved
doi: 10.31254/phyto.2025.14306

Anupsingh Vijaysingh Thakur
Department of Molecular Biology and Genetic Engineering, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar- 263145, Uttarakhand, India

Sonu Ambwani
Department of Molecular Biology and Genetic Engineering, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar- 263145, Uttarakhand, India

Tanuj Kumar Ambwani
Department of Veterinary Physiology and Biochemistry, College of Veterinary and Animal Sciences, G.B. Pant University of Agriculture and Technology, Pantnagar- 263145, Uttarakhand, India

Sudhir Kumar
Department of Veterinary Physiology and Biochemistry, College of Veterinary and Animal Sciences, G.B. Pant University of Agriculture and Technology, Pantnagar- 263145, Uttarakhand, India

Correspondence:

Dr. Sonu Ambwani
Department of Molecular Biology and Genetic Engineering, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar- 263145, Uttarakhand, India
Email: sonuambwani@yahoo.co.in

Assessment of immunomodulatory properties of *Acacia catechu* leaf extract employing chicken lymphocyte culture system

Anupsingh Vijaysingh Thakur, Sonu Ambwani, Tanuj Kumar Ambwani, Sudhir Kumar

ABSTRACT

Background: Medicinal plants have long been integral to traditional healthcare systems, offering a rich source of bioactive compounds beneficial for both human and animal health. Their accessibility, low toxicity, and therapeutic potential make them valuable alternatives or complements to modern pharmaceuticals. *Acacia catechu* (L.f.) Willd. (*Senegalia catechu*) (family: Fabaceae), a widely used plant in traditional Ayurvedic and folk medicine, has garnered attention for its diverse pharmacological properties. Various parts of the plant particularly the bark, heartwood, and leaves are traditionally employed for treating ailments such as diarrhea, cough, skin diseases, and inflammation. Recent studies have highlighted the immunomodulatory potential of *Acacia catechu*. **Objective:** The fifty percent hydromethanolic leaf extract of *Acacia catechu* (ACLE) was prepared and evaluated for its *in vitro* immunomodulatory potential using animal cell-culture model of avian lymphocytes. **Materials and Methods:** In this study, lymphocytes were isolated from peripheral blood of healthy chickens using density gradient centrifugation. The isolated cells were cultured and treated with varying concentrations of ACLE under standard *in vitro* conditions. Mitogens such as Concanavalin A (ConA) and lipopolysaccharide (LPS) were used to stimulate T and B cell proliferation, respectively. Cell viability and proliferation were assessed using the MTT assay. **Results:** MTT assay revealed a dose-dependent enhancement of lymphocyte proliferation in the presence of the extract. Considering the outcome of MTT assay, 0.3 mg/mL dose was chosen as the maximum non-cytotoxic dose (MNCD) for further immunological evaluations, as it maintained cell viability without inducing cytotoxic effects. Lymphocytes proliferation assay (LPA) demonstrated the immunomodulating potential under mitogenic stimulation. **Conclusion:** *In vitro* experiments using chicken lymphocytes have demonstrated significant immunomodulatory activity. These findings underscore the potential of *Acacia catechu* as a natural immunomodulator in poultry health management, contributing to improved disease resistance and reduced reliance on synthetic antibiotics. Further exploration and validation of such traditional botanicals may support the development of sustainable and effective herbal therapeutics for both veterinary and human applications.

Keywords: *Acacia catechu*, *Senegalia catechu*, Immunomodulation, Chicken lymphocytes, Lymphocytes proliferation assay.

INTRODUCTION

Medicinal plants have long been integral to maintaining the health of both humans and animals, with their benefits extending across multiple sectors, including medicine, agriculture, food, and cosmetics. Historical records from ancient civilizations such as those in India, China, Mesopotamia, and Egypt reveal extensive use of botanical remedies for the prevention and treatment of various diseases^[1,2]. Prior to the advent of modern synthetic pharmaceuticals, these plants formed the foundation of traditional healthcare systems, offering natural treatments rich in biologically active substances. Advances in fields like phytochemistry and pharmacognosy have provided scientific validation for many traditional practices, linking specific plant-derived compounds—such as alkaloids, flavonoids, terpenoids, and tannins—to therapeutic effects^[3,4]. In recent decades, there has been a marked revival of interest in herbal medicine, often referred to as a “herbal renaissance,” driven by a preference for natural remedies, awareness of adverse effects associated with synthetic drugs, and renewed appreciation for traditional ethnomedical knowledge^[4].

Acacia catechu (L.f.) Willd (*Senegalia catechu*) (family: Fabaceae), commonly referred to as khair, black cutch, catechu, or cachou, is a deciduous tree native to India and widely distributed across other parts of Asia, including Nepal, Bangladesh, Pakistan, Myanmar, Thailand, and Indonesia, as well as regions of East Africa^[5]. Botanically, *Acacia catechu* is a medium-sized tree ranging from 5 to 15 meters in height. Its straight stem is grayish-brown in color, and the bark is dark brown with reddish

inner layers that exfoliate in narrow strips [6]. The leaves are bipinnately compound, comprising 10–30 pairs of pinnae, each bearing 20–50 pairs of small leaflets. The plant also features short, hooked spines and axillary pedunculate spike inflorescences. A decoction made from the heartwood is traditionally consumed as a health beverage in southern India, believed to purify the blood, enhance skin health, and strengthen the immune system [7]. In various traditional medicinal systems, *Acacia catechu* has been employed for its antimicrobial, anti-inflammatory, antidiarrheal, astringent, antimycotic, and coagulant properties [8,9,10,11]. It is also known for its vermifuge action and has been traditionally used to manage conditions such as diabetes, obesity, and to promote wound healing and dental hygiene [8,12,13,14]. The heartwood extract, commonly known as black catechu, is used to treat respiratory conditions such as asthma and bronchitis, as well as gastrointestinal disorders like colic and diarrhea [15]. It is also applied in the treatment of skin ailments, boils, ulcers, and stomatitis [14]. Pharmacological studies have confirmed the bark's anthelmintic, antipyretic, and anti-inflammatory effects [16,17]. Recent research has highlighted the immunomodulatory potential of the bark extract of *Acacia catechu*, suggesting its role in enhancing host defense mechanisms [7,18]. In traditional veterinary medicine, the bark decoction (500 ml) of *Senegalia modesta* (*Accacia modesta*) is mixed with 200 g butter oil and is given orally twice a day for 5 days to cows and buffaloes for easy parturition and for expelling the placenta [19]. Furthermore, in Chinese traditional medicine, a formulation known as “Ercha,” derived from the heartwood extract of *Acacia catechu*, is widely used in the treatment of dysentery, Ulcerative colitis [20]. Extracts of the heartwood showed positive results for tannins, terpenoids, triterpenoids, alkaloids, ascorbic acid, and carbohydrates, while leaf extracts also exhibited tannin and saponins [21,22,23,24].

The leaves of *Acacia catechu* have been reported to contain significant structural and nutritional components, including hemicellulose, lignin, cellulose, and essential minerals. Phytochemical analysis of its ethanolic leaf extract revealed the presence of various flavonoids such as quercetin, poriferasterol- β -D-glucoside, quercetin-3-O-arabinofuranoside, quercetin-3-O-rhamnoside, and quercetin-3-O-galactoside [25,26]. Considering above, this study aimed to evaluate the *in vitro* immunomodulatory activity of a 50% hydro-methanolic extract derived from *Acacia catechu* leaves (ACLE) using a chicken lymphocyte cell culture model.

MATERIALS AND METHODS

Collection and Preparation of the Plant Extract

Fresh and taxonomically authenticated leaves of *Acacia catechu* were harvested from the Agroforestry Research Centre (AFRC) at Govind Ballabh Pant University of Agriculture and Technology, Pantnagar, Uttarakhand, India. A 50% hydro-methanolic extract of the leaves (ACLE) was prepared following the previously described method [23,27]. To maintain consistency and sterility across all downstream applications, only cell culture-grade reagents and solvents were used throughout the experimental procedures.

Evaluation of *In vitro* Immunomodulatory Properties of ACLE

Isolation of Chicken Lymphocytes

Peripheral blood samples were obtained from healthy broiler chickens aged between 4–6 weeks, procured from a local poultry processing facility. Samples were immediately transferred to the laboratory in sterile condition. Lymphocytes were isolated under aseptic conditions using standard density gradient separation techniques to ensure high cell viability. More than 95% cell viability was confirmed via the trypan blue exclusion assay, indicating the integrity of the isolated lymphocyte population. The study is *in vitro* in nature involving blood samples collected from the slaughtered healthy birds therefore do not require Institutional Animal Ethics Permission.

Determination of Maximum Non-Cytotoxic Dose (MNCD) and Lymphocyte Proliferation Assay (LPA)

The cytotoxicity of *Acacia catechu* leaf extract (ACLE) was assessed using the MTT assay, which measures mitochondrial-dependent cellular activity [28,29]. Chicken lymphocytes were seeded in 96-well flat-bottom plates at a density of 1×10^6 cells/mL and exposed to a gradient of ACLE concentrations ranging from 0.001 to 1.6 mg/mL in triplicates following 68 hours of incubation at 40°C in a humidified 5% CO₂ incubator. Subsequently, 20 μ L of MTT solution (5 mg/mL) was added to each well, and incubation was continued for 4 hours in the dark. Formazan crystals formed by viable cells were dissolved by adding 200 μ L of DMSO per well. Absorbance was recorded at 570 nm using an ELISA microplate reader. Cell viability was expressed as a percentage relative to the untreated control group.

To assess the immunomodulatory potential of ACLE, a lymphocyte proliferation assay (LPA) was performed following established protocols [30]. Chicken lymphocytes (1×10^6 cells/mL) were seeded in 96-well tissue culture plates with or without mitogenic stimulation. The mitogens used included Concanavalin A (ConA) and *E. coli* lipopolysaccharide (LPS), each at a final concentration of 5 μ g/mL. The MNCD of ACLE was added to the respective wells, and all treatments were conducted in triplicate to ensure reproducibility and statistical reliability.

Statistical Analysis

Experimental data were statistically analyzed using one-way analysis of variance (ANOVA) with Least Significant Difference (LSD) post-hoc test to compare control and treated groups. Results are presented as mean \pm standard error (SE). Pearson correlation analysis was employed to examine associations between variables. Statistical significance was defined as $p < 0.05$. All analyses were performed using SPSS statistical software.

RESULTS

The Plant Extract (ACLE)

The percent yield of fifty percent hydromethanolic plant extract *i.e.*, ACLE obtained was 10.46% that was prepared using dried leaf powder of *Acacia catechu* (Figure 1).

Evaluation of Immunomodulatory properties of ACLE

To determine the maximum non-cytotoxic dose (MNCD) suitable for *in vitro* immunological studies, chicken lymphocytes were exposed to a range of concentrations of the 50% hydro-methanolic extract of *Acacia catechu* leaves (ACLE). The viability of the cells was assessed using the MTT assay, which revealed a dose-dependent cytotoxic effect. Higher concentrations of the extract, particularly between 0.5 mg/mL and 1.6 mg/mL, were associated with a notable decline in cell viability. At the highest tested concentration (1.6 mg/mL), cell viability decreased significantly, showing 32.58% cytotoxicity. In contrast, lower concentrations till 0.3 mg/mL maintained 100% viability of the lymphocytes. Based on these observations, 0.3 mg/mL the highest concentration that did not impair cell viability was selected as the MNCD for subsequent immunological assays.

When lymphocytes were treated with the MNCD of ACLE, a slight increase of 5.33% in overall lymphocyte proliferation was observed compared to the untreated control, indicating potential immunostimulatory activity. Notably, treatment with ACLE enhanced lipopolysaccharide (LPS)-stimulated lymphocyte proliferation by 14.75%, suggestive of augmented B-cell activation and blastogenesis. Conversely, a slight reduction in Concanavalin A (ConA)-stimulated proliferation was noted, with cell viability maintained at 94.15%, indicating a marginal suppression of T-cell responsiveness. These findings suggest a differential modulatory effect of *Acacia catechu*

leaf extract on T and B lymphocytes, with a stronger stimulatory influence on B-cell activity.

Table 1: Immunomodulatory potential due to *in vitro* exposure of CTE in chicken lymphocytes

Treatment	Un-stimulated		LPS stimulated		ConA stimulated	
	Control	ACLE	Control	ACLE	Control	ACLE
% Viability*	100	105.34	100	114.75	100	94.15
Cd value at 1 %	0.046622	4.577562*	0.122522	4.596257*	0.054151	11.65769**
Cd value at 5 %	0.033269		0.080395		0.038642	



Figure 1: *Acacia catechu* plant material and extract. (a) *Acacia catechu*, (b) Dried leaf powder of *Acacia catechu*, (c) Fifty percent hydromethanolic leaf extract of *Acacia catechu*

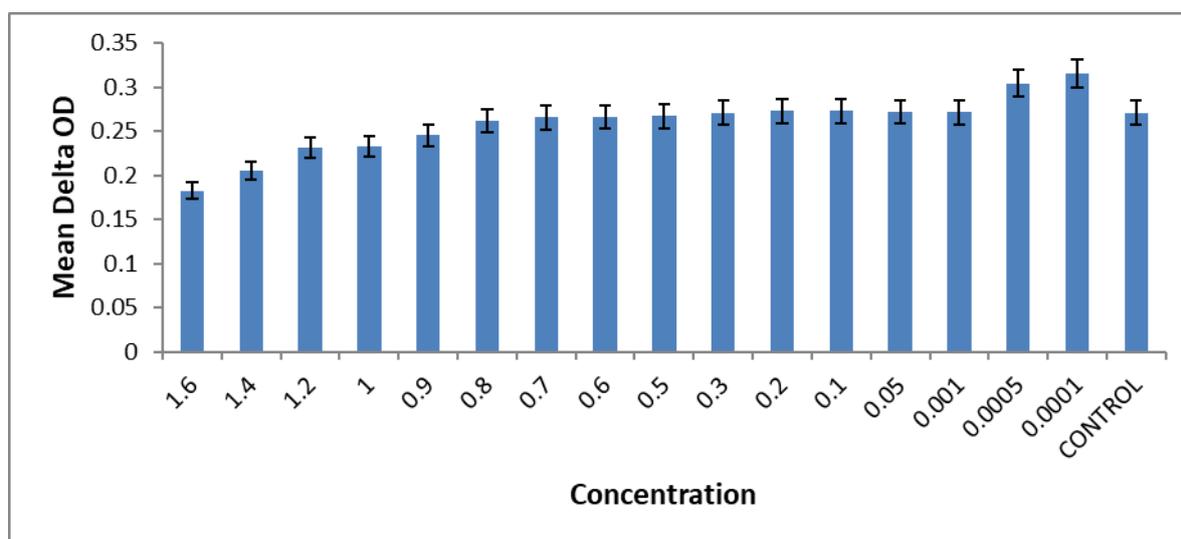


Figure 2: Non-cytotoxic dose of *Acacia catechu* extract by MTT cytotoxicity assay

DISCUSSION

The global resurgence in the use of herbal medicine over recent decades is driven by a growing interest in plant-based therapeutic agents with fewer side effects and broader biological activities. *Acacia catechu* (commonly known as Black Cutch or Black Kutch, khair or kath) is a well-recognized medicinal plant employed in both traditional and contemporary medical systems [31,32]. This species has been used extensively in Ayurvedic and Unani practices for the treatment of various ailments, including diarrhea, cough, ulcers, and skin infections [33]. Phytochemical Composition *Acacia catechu* is notably rich in secondary metabolites such as flavonoids, tannins, polyphenols, and terpenoids. Almost all parts of the plant—including the heartwood, bark, leaves, roots, and flowers—harbor biologically active compounds; however, the heartwood and leaves are particularly pharmacologically significant [14,21,22,24].

The heartwood of *Acacia catechu* is a source of key phytochemicals including catechin, epicatechin, epigallocatechin, catechin tetramers, taxifolin, kaempferol, quercetin, quercitrin, phlobatannin, gossypetin, protocatechuic acid, phloroglucin, and various glycosides and sugars such as L-arabinose and D-galactose [10,21,22,25]. In addition to flavonoids and tannins, GC-MS analysis of leaf extracts has identified terpene compounds such as camphor (76.40%) and phytol (27.56%), which contribute to the plant's antimicrobial, DNA protective and antifungal properties [10,34,35,36]. Further GC-MS profiling revealed the presence of Caprylic acid methyl ester (19.77%), Lauric acid methyl ester (27.42%), 2-ethyl-3-methyl-1-butene (42.09%), and Myristic acid methyl ester (10.72%), highlighting the chemical diversity of the species [23].

The therapeutic potential of *Acacia catechu* is strongly correlated with its antioxidant activity, mainly due to its high phenolic and flavonoid

content. Quercetin, a prominent phenolic flavonoid, plays a significant role in neutralizing free radicals and contributing to the plant's antioxidative defense mechanism [23,37,38]. Taxifolin, another key flavonoid, has demonstrated a broad range of biological activities including antifungal, antiviral, anti-inflammatory, and antioxidant effects [26,38,39]. Taxifolin isolated from the leaf extract has been found to exert potent antibacterial activity against *Streptococcus mutans* and *Lactobacillus acidophilus*, with inhibition zones of 23 mm and 14.5 mm, respectively at 2.5 mg/mL, underscoring its efficacy as an antimicrobial agent [25]. Saha *et al.* [17] reported substantial antioxidant activity in the leaf extract of *Acacia catechu*, suggesting that the pronounced scavenging effects against various reactive oxygen species were due to the rich flavonoid and phenolic content. *In vitro* assays using rat liver post-mitochondrial supernatant systems showed that the extracts of *Acacia catechu* significantly inhibited both the initiation and propagation of lipid peroxidation [40]. The extract also demonstrated potent hydrogen peroxide scavenging capacity [23]. In a related study, Hédi *et al.* [41] observed that *Acacia salicina*, a species of the same genus, also exhibits anti-genotoxic and antioxidant activity, attributed primarily to its sterol and phenolic contents. They noted that the antioxidant potential of leaf extracts was relatively lower compared to bark and heartwood, a finding that aligns with observations in *Acacia catechu*. Similarly, Stohs & Bagchi [16] further substantiated its antioxidant, anti-inflammatory and chemoprotective benefits—key to reducing cellular damage. Our findings displayed *in vitro* immunomodulatory properties of ACLE in cell culture model system of lymphocytes that can be attributed to the presence of various phytochemicals present in the extract. Ismail & Asad [18] and Sunil *et al.* [7] provided compelling *in vitro* and *in vivo* evidence that *Acacia catechu* extracts can modulate both innate and adaptive immunity. The plant also exhibits a variety of other pharmacological effects including antipyretic, hypoglycemic [17,42], antidiarrheal, antiulcer, and hepatoprotective properties [8,42]. Saha *et al.* [17] elucidate the plant's antioxidant capabilities, which may counteract oxidative stress implicated in cognitive disorders. These properties enhance the plant's value as a traditional medicine source.

CONCLUSION

On the basis of the outcome of this study, it can be concluded that the fifty percent hydro-methanolic leaf extract of *Acacia catechu* exhibited significant *in vitro* immunomodulating properties in chicken lymphocytes. There was increase in lymphocytes proliferation under LPS stimulation while under ConA stimulation marked decrease was observed. The wide-ranging pharmacological properties of *Acacia catechu* are largely due to its complex and diverse phytochemical profile. Its antioxidant, antimicrobial, immunomodulatory, and hepatoprotective properties make it a valuable plant for the development of herbal therapeutics. While traditional uses support its medicinal relevance, further advanced analytical and clinical studies are essential to fully explore and validate the therapeutic potential of this important medicinal species.

Acknowledgments

Authors are thankful to the Director, AFRC, G.B.P.U.A. &T., Pantnagar, for providing the plant material. The facilities provided by Director Experiment Station; Dean, Veterinary & Animal Sciences, GBPUA&T, Pantnagar, to carry out the present study, are duly acknowledged. Master's thesis grant provided to A.V. Thakur by DBT, New Delhi, India is duly acknowledged.

Conflict of interest

The authors declared no conflict of interest.

Financial Support

None declared.

ORCID ID

Sonu Ambwani: <https://orcid.org/0000-0002-6202-3115>

Tanuj Kumar Ambwani: <https://orcid.org/0000-0001-6871-135X>

Sudhir Kumar: <https://orcid.org/0009-0002-8236-9364>

REFERENCES

1. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Front Pharmacol.* 2014;4:177.
2. Ambwani S, Tandon R, Ambwani TK, Malik YS. Current knowledge on nanodelivery systems and their beneficial applications in enhancing the efficacy of herbal drugs. *J Exp Biol Agric Sci.* 2018;6(1):87–107.
3. Ambwani S, Tandon R, Ambwani TK. Metal nano delivery systems for improved efficacy of herbal drugs. *Biosci Biotechnol Res Asia.* 2019;16(2):251–261.
4. Pandey Y, Ambwani S. Nano metal-based herbal theranostics for cancer management: Coalescing nature's boon with nanotechnological advancement. *Curr Pharm Biotechnol.* 2022;23(1):30–46.
5. Lakshmi T, Roy A, Geetha R. *Acacia catechu* Willd—A gift from Ayurveda to mankind: a review. *Res J Pharm Res.* 2011;5(2):273–293.
6. Kirtikar KR, Basu BD. *Acacia* Tourn. Linn. In: Blatter E, Caius JF, Mhaskar K, editors. *Indian Medicinal Plants*. 2nd ed. Dehradun: Int Book Distributors; 1935. p. 919–935.
7. Sunil MA, Sunitha VS, Radhakrishnan EK, Jyothis M. Immunomodulatory activities of *Acacia catechu*, a traditional thirst quencher of South India. *J Ayurveda Integr Med.* 2019;10(3):185–191.
8. Ray D, Sharatchandra KH, Thokchom IS. Antipyretic, antidiarrhoeal, hypoglycaemic and hepatoprotective activities of ethyl acetate extract of *Acacia catechu* Willd in albino rats. *Indian J Pharmacol.* 2006;38(6):408–413.
9. Nagaraja TG, Sarang SV, Jambhale DC. Evaluation of antimycotic activity of *Acacia catechu* Willd. *Mimosaceae.* *J Biopestic.* 2008;1(2):197–198.
10. Ramesh B, Jayabharathi V. Phytochemical screening, HPTLC and GC-MS profile of *Acacia catechu* (L.f.) Willd hydroethanolic leaf extract. *Int J Curr Microbiol App Sci.* 2017;6(1):82–94.
11. Ahamad ST, Lakshmi T, Rajeshkumar S, Roy A, Gurunadhan D, Geetha R. Antibacterial activity of taxifolin isolated from *Acacia catechu* leaf extract—an *in vitro* study. *Indian J Public Health Res Dev.* 2019;10(11):3540.
12. Singh KN, Mittal RK, Barthwal KC. Hypoglycemic activity of *Acacia catechu*, *Acacia suma*, and *Albizia odoratissima* seed diets in normal albino rats. *Indian J Med Res.* 1976;64(5):754–757.
13. Jarald E, Joshi SB, Jain DC. Biochemical study on the hypoglycaemic effects of extract and fraction of *Acacia catechu* Willd in alloxan-induced diabetic rats. *Int J Diabetes Metab.* 2009;17:63–69.
14. Kumari M, Radha, Kumar M, Zhang B, Amarowicz R, et al. *Acacia catechu* (L.f.) Willd.: a review on bioactive compounds and their health promoting functionalities. *Plants (Basel).* 2022;11(22):3091.
15. Karwani G, Singhvi I, Gupta S, Kapadiya N, Sisodia SSI. Antisecretory and antiulcer activity of *Acacia catechu* against indomethacin plus pyloric ligation induced gastric ulcers in rats. *J Cell Tissue Res.* 2011;11(1):2567–2571.
16. Stohs SJ, Bagchi D. Antioxidant, anti-inflammatory and chemoprotective properties of *Acacia catechu* heartwood extracts. *Phytother Res.* 2015;29(6):818–824.
17. Saha MR, Dey P, Begum S, De B, Chaudhuri TK, Sarker DD, et al. Effect of *Acacia catechu* (L.f.) Willd. on oxidative stress with possible implications in alleviating

- selected cognitive disorders. PLoS One. 2016;11(3):e0150574.
18. Ismail S, Asad M. Immunomodulatory activity of *Acacia catechu*. Indian J Physiol Pharmacol. 2009;53(1):25–33.
 19. Rehman S, Iqbal Z, Qureshi R. Ethnoveterinary practices of medicinal plants among tribes of tribal district of North Waziristan, Khyber Pakhtunkhwa, Pakistan. Front Vet Sci. 2022;9:815294.
 20. Zhang S, Zhao L, Shen H. International clinical practice guideline on the use of traditional Chinese medicine for ulcerative colitis. Phytoter Res. 2023;38(2):970–999.
 21. Shen D, Wu Q, Wang M, Yang Y, Lavoie EJ, Simon JE. Determination of the predominant catechins in *Acacia catechu* by LC/ESI-MS. J Agric Food Chem. 2006;54:3219–3224.
 22. Li XC, Liu C, Yang LX, Chen RY. Phenolic compounds from the aqueous extract of *Acacia catechu*. J Asian Nat Prod Res. 2011;13:826–830.
 23. Thakur AV, Ambwani S, Ambwani TK. Preliminary phytochemical screening and GC-MS analysis of leaf extract of *Acacia catechu* (L.f.) Willd. Int J Herbal Med. 2018;6(2):81–85.
 24. Aryal B, Adhikari B, Aryal N, et al. LC-HRMS profiling and antidiabetic, antioxidant, and antibacterial activities of *Acacia catechu* (L.f.) Willd. Biomed Res Int. 2021;2021:7588711.
 25. Negi BS, Dave BP. In vitro antimicrobial activity of *Acacia catechu* and its phytochemical analysis. Indian J Microbiol. 2010;50(4):369–374.
 26. Ezhil I, Lakshmi T. Antibacterial efficacy of epicatechin and rutin from *Acacia catechu* leaf extract against *Enterococcus faecalis* and *Streptococcus mutans*: an *in vitro* study. J Adv Pharm Edu Res. 2017;7(1):22–24.
 27. Thakur AV, Ambwani S, Ambwani TK, Ahmad AH, Rawat DS. Evaluation of phytochemicals in the leaf extract of *Clitoria ternatea* Willd. through GC-MS analysis. Trop Plant Res. 2018;5(2):200–206.
 28. Kakade DP, Arora S, Ambwani S. Anti-proliferative effect of silver nanoparticles in HeLa cells due to enhanced oxidative stress. Res J Biotechnol. 2018;13(2):68–74.
 29. Osman SOM, Paliwal L, Ambwani S, Ambwani TK. Comparative analysis of *Albizia lebbek* and *Lawsonia inermis*: phytochemical estimation and their immunomodulatory effects. J Phytopharmacol. 2024;13(6):464–469.
 30. Ambwani S, Dolma R, Sharma R. Modulation of inflammatory and oxidative stress biomarkers due to dexamethasone exposure in chicken splenocytes. Vet Immunol Immunopathol. 2023;262:110632.
 31. Singh KN, Lal B. Notes on traditional uses of khair (*Acacia catechu* Willd.) by inhabitants of Shivalik range in Western Himalaya. Ethnobot Leaflets. 2006;10:109–112.
 32. Bhattarai R, Sharma P, Wagle B. Revision and compilation of health management plan of khair (*Acacia catechu*). Grassroots J Nat Resour. 2020;3(1):15–28.
 33. Sharma KR, Kalauni SK, Awale S, Pokharel YR. *In vitro* free radical scavenging activity of methanol extracts of some selected medicinal plants of Nepal. Austin J Biotechnol Bioeng. 2015;2(1):1035.
 34. Patel JD, Kumar V, Bhatt SA. Antimicrobial screening and phytochemical analysis of the resin part of *Acacia catechu*. Pharm Biol. 2009;47:34–37.
 35. Hazra B, Sarkar R, Biswas S, Mandal N. The antioxidant, iron chelating and DNA protective properties of ‘katha’ heartwood extract of *Acacia catechu*. J Complement Integr Med. 2010;7(1).
 36. Silva E, Fernandes S, Bacelar E, Sampaio A. Antimicrobial activity of aqueous, ethanolic and methanolic leaf extracts from *Acacia* spp. and *Eucalyptus nicholii*. Afr J Tradit Complement Altern Med. 2016;13(6):130–134.
 37. Jayasekhar P, Mohanan PV, Rathinam K. Hepatoprotective activity of ethyl acetate extract of *Acacia catechu*. Indian J Pharmacol. 1997;29:426–428.
 38. Patil A, Modak M. Comparative evaluation of oxidative stress modulating and DNA protective activities of aqueous and methanolic extracts of *Acacia catechu*. Medicines. 2017;4(3):65.
 39. Lakshmi T, Rajendran R, Madhusudhanan N. Chromatographic fingerprint analysis of *Acacia catechu* ethanolic leaf extract by HPTLC technique. Int J Drug Dev Res. 2012;4:180–185.
 40. Yadav AS, Bhatnagar D. Inhibition of iron induced lipid peroxidation and antioxidant activity of Indian spices and *Acacia in vitro*. Plant Foods Hum Nutr. 2009;65:18–24.
 41. Hédi BM, Jihed B, Inks B. Antigenotoxic activities of crude extracts from *Acacia salicina* leaves. Environ Mol Mutagen. 2007;48:58–66.
 42. Patil A, Latake P, Kumbhar S, Chougule N. Pharmacological activities of *Acacia catechu*. Res J Pharmacogn Phytochem. 2024;16(4):263–269.

HOW TO CITE THIS ARTICLE

Thakur AV, Ambwani S, Ambwani TK, Kumar S. Assessment of immunomodulatory properties of *Acacia catechu* leaf extract employing chicken lymphocyte culture system. J Phytopharmacol 2025; 14(3):165-169. doi: 10.31254/phyto.2025.14306

Creative Commons (CC) License-

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY 4.0) license. This license permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. (<http://creativecommons.org/licenses/by/4.0/>).