

The Journal of Phytopharmacology

(Pharmacognosy and phytomedicine Research)

Research Article

ISSN 2320-480X
JPHYTO 2017; 6(5): 307-310
September- October
Received: 07-10-2017
Accepted: 15-11-2017
© 2017, All rights reserved

Refaz Ahmad Dar

Division of Biotechnology, Indian Institute of Integrative Medicine (CSIR), Sanatnagar, Srinagar, Jammu & Kashmir-190005, India

Iram Saba

Division of Biotechnology, Indian Institute of Integrative Medicine (CSIR), Sanatnagar, Srinagar, Jammu & Kashmir-190005, India

Mohd Shahnawaz

Division of Plant Biotechnology, Indian Institute of Integrative Medicine (CSIR), Jammu - Tawi, Jammu & Kashmir-180001, India

Parvaiz Hassan Qazi

Division of Biotechnology, Indian Institute of Integrative Medicine (CSIR), Sanatnagar, Srinagar, Jammu & Kashmir-190005, India

Inshad Ali Khan

Division of Clinical Microbiology, Indian Institute of Integrative Medicine (CSIR), Jammu - Tawi, Jammu & Kashmir-180001, India

Correspondence:

Parvaiz Hassan Qazi
Division of Biotechnology, Indian Institute of Integrative Medicine (CSIR), Sanatnagar, Srinagar, Jammu & Kashmir-190005, India
Email: qphassan[at]iiim.ac.in

Antimicrobial potential of fungal endophytes from selected high value medicinal plants of the Kashmir valley – India

Refaz Ahmad Dar, Iram Saba, Mohd Shahnawaz, Parvaiz Hassan Qazi*, Inshad Ali Khan

ABSTRACT

The purpose of this work was to evaluate the antimicrobial potential of endophytic fungi isolated from different high value medicinal plants of Kashmir valley. Evaluation of some endophytes has been carried for their possible antimicrobial activity from various parts of medicinal plants belonging to Kashmir valley (India). A total of twenty-eight fungal endophytes were isolated from the different parts of selected medicinal plants. Dichloromethane (DCM) extracts of all the morphologically different endophytes were prepared and subsequently checked for antimicrobial activities. Eight isolates showed good activity against gram positive bacteria with two isolates showing promising activity with MIC in the range of 0.5 – 1µg/ml. All the isolated endophytic extracts were completely devoid of antifungal activity. The seven-active endophytic fungal cultures were identified by ITS4 and ITS5 gene sequencing.

Keywords: Endophyte, Antimicrobial, MIC.

INTRODUCTION

Recently endophytes are viewed as outstanding source of secondary metabolites and bioactive antimicrobial natural products. Endophytic fungi are of biotechnological interest due to their potential use as genetic vectors, metabolites^[1] and biological control agents^[2]. In the last few decades, scientists have realized that plants are as a reservoir of unexplored numbers of potential microorganisms known as endophytes. These endophytic microbes can produce secondary metabolites such as enzymes growth hormones, antimicrobial, anti-fungal or as anticancer substances^[3, 4]. These secondary metabolites were proved potential drugs for treatment of newly developing disease in humans, plants and animals^[5]. Due to host-endophyte co-evolution, some plants that produce bioactive natural products have associated endophytes that produce the same natural products^[6]. From the microbial sources the isolation of bioactive compounds is easier and more economical for large-scale production than plant sources. The production of metabolites by microorganisms is known and explored, because the major of antibiotics are produced by fungi and bacteria^[7]. Antimicrobial activities have been demonstrated in a variety of metabolites biosynthesized by plant endophytes^[8]. There is an increasing effort to characterize and identify the endophytic fungi isolated from the medicinal plants. Azevedo *et al.*,^[9] reported that some medicinal properties of plants may be related to endophytic fungi hosted by these plants.

Endophytes have been intensively studied in several unexplored environments around the world. A little work in this line has been done from this state so far (which is endowed with a rich biodiversity of plant species and it has a long tradition of herbal medicine and the need for the isolation of endophytic microbes from these is the need of an hour. This led us for the isolation and exploitation of the bioactive potential of the endophytic microbes from Kashmir valley.

In the present study an attempt has been made to isolate the endophytic fungi from the selected high value medicinal plants of Kashmir valley and assessed their antimicrobial potentials at *in vitro* level. These endophytic fungi may be the source of the potential antimicrobial drugs.

MATERIALS AND METHODS

Collection and identification of plant material

High value medicinal plants were collected from two localities, Sonamarg, Kashmir valley, J & K, India and from the field of CSIR-Indian Institute of Integrative Medicine (IIIM), Sanantnagar, J & K, India. Four types of plants in 5 replicates were collected randomly from each locality by individual and

group visits (*Rheum emodi* wall. ex Meissn and *Hypericum perforatum* L. from Sonamarg and *Diocoria deltodia* Wall. and *Artemisia annua* L. from the fields of IIIM Sanantnager).

Isolation and purification of different endophytic strains

From each plant type, endophytic fungi were isolated as per Arnold *et al.*,^[10] with some modifications. Plant materials (leaf, stem and root) were thoroughly washed with running tap water followed by immersion in 70% EtOH for 1 min and in NaOCl (2.5% - 5.25%) for 3 min, drained and immersed in 70% EtOH again for 30 sec. Finally, the samples were rinsed with sterile distilled water and cut under sterile conditions into small pieces (2–3 cm). The internal tissues were cut into smaller pieces of 0.5 to 1 cm and plated on different media such as water agar, potato dextrose agar and rose bengal agar (Difco) supplemented with streptomycin sulphate (250 µg/ml, Sigma) at 28°C for 2 -3 weeks. The fungi coming out of the plant tissues were taken and grown on PDA plates free of antibiotics as a pure culture. The pure cultures were also preserved in glycerol and submitted at IIIM microbial repository. The morphological identification of the endophytic strains were done by preparing microscopic slides stained with lactophenol cotton blue as per the key of Vainio *et al.*,^[11] and were examined under light microscope (Olympus, USA).

Extraction of the fungal endophytes

The endophytic fungal strains were grown on potato dextrose broth at 180 rpm, 28 °C for 10-15 days in an incubator shaker (New Brunswick, USA). The grown culture of each endophyte was homogenized with 10% methanol under ultra sonicator. The fermented broth was then extracted with Dichloromethane three times after which Na₂SO₄ (40µg/ml, Merck) was added to further remove the aqueous layer within the mixture. The mixture was then transferred to a round bottom flask and dried using a rotary evaporator and weighed to constitute the crude broth extract.

Screening of the endophytic fungal extracts for antimicrobial activity

For the screening the antimicrobial activity the following media were used such as Muller Hinton Agar, Muller Hinton Broth, Sabroude's Dextrose broth and Sabroude's Dextrose agar (Becton-Dickinson, Cockeysville, MD, USA; DIFCO laboratories) and the extracts were tested against following bacteria: Gram positive *Staphylococcus aureus* ATCC 29213, *Methicillin resistant Staphylococcus aureus* ATCC 15187, *Vancomycin resistant enterococcus faecalis* (VRE), Gram negative *Escherichia coli* ATCC 25292, *Pseudomonas aeruginosa* ATCC 27853. Fungal strains *Candida albicans* ATCC 90028, *C. albicans* V-0-191 and *Aspergillus niger* ATCC 16404 and *A. fumigates*. All the cultures were procured from the American Type Cultures Collection (Manassas, VA, USA). Bacterial cultures were maintained on Tryptone soya agar, fungal cultures were maintained on Sabroude's Dextrose agar, and stored at -70 °C containing 50% glycerol. Antibiotic were obtained from sigma –Aldrich, Ciprofloxacin and Amphotericin- B was used as a standard antibacterial and antifungal agent for this study. The concentrations of the stock solutions were 1mg/ml.

Minimum inhibitory concentration (MIC) assay

MIC was determined as per the guidelines of Clinical and Laboratory Standards Institute^[12]. Bacterial and fungal suspensions were

prepared by suspending 18 h grown bacterial culture and 24h fungal culture in sterile normal saline. The turbidity of the bacterial suspension was adjusted to 0.5 McFarland standards (equivalent to 1.5 x 10⁸ CFU/ml) and 1.0 McFarland standards (equivalent to 1.5x10⁸ colony forming units (CFU)/ml) for fungal stains at wavelength 625nm. The extract stock solutions were prepared in 100% dimethyl sulfoxide (DMSO; Merck, Mumbai India) and 2-fold serial dilutions were prepared in Mueller Hinton Broth for bacteria and Sabroude's Dextrose broth for fungal in 100 µl volume in 96-well U bottom microtitre plates (Tarson, Mumbai, India). The above mentioned bacterial and fungal suspension were further diluted in the MHB and SDB and 100 µl volume of this diluted inoculum was added to each well of the plate resulting in the final inoculum of 5x10⁵ CFU/mL in the well and the final concentration of extracts ranged from 0.5 to 256 µg/ml. Ciprofloxacin and Amphotericin B was used as standard Antibacterial and antifungal agent for this study at a concentration ranged from 0.03-16 µg/ml. The bacterial plates were incubated at 37°C for 18 h and fungal plates at 28°C for 42 h and were visually read for the absence or presence of turbidity. The minimum concentration of the compound concentration showing no turbidity was recorded as MIC.

Identification of the potent fungal endophytes

Identification of the fungal endophytes was based on their internal transcribed spacer ribosomal DNA (ITSrDNA) sequences. A pair of primers ITS4 (sequence: 5'-TCC GTA GGT GAA CCT GCG G-3') and ITS5 (5'-TCC TCC GCT TAT TGA TAT GC-3') was used for ITS-rDNA amplification, using the polymerase chain reaction (PCR). The consensus sequences were subjected to BLAST searches of the NCBI GenBank database (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) for identification and to find out the sequence homology with closely related organisms. All the endophytic fungal ITS-rDNA sequences were deposited in GenBank and the accession number was assigned to each sequence.

RESULTS

Isolation of the fungal endophytes

From all the four selected medicinal plants total 28 fungal endophytes have been isolated. Based on the morphology of the endophytes 14 different endophytes have been selected for antimicrobial activities. The selected endophytes are IIIM1, IIIM2, IIIM3, IIIM4, IIIM5, IIIM6, IIIM7, IIIM8, IIIM9, IIIM10, IIIM11, IIIM12, IIIM13, and IIIM14.

Molecular identification and characterization of endophytic fungi

The bioactive strains were identified up to the genus level using ITS1-5.8S-ITS2 approach. Among the identified fungi, *Pichia kudriavzevii* (query cover 100%, max identity 99%), *Fusarium oxysporum* (query cover 97%, max identity 99%), *Mucor circinelloide* (query cover 99%, max identity 99%), *Mucor circinelloide* (query cover 99%, max identity 87%), *Trametes versicolor* (query cover 94%, max identity 95%), *Polyporales sp.* (query coverage 100%, max identity 99%), *Bjerkandera adusta* (query coverage 98%, max identity 99%), *Fusarium tricinctum* (query coverage 100%, max identity 100%). The sequences was submitted to NCBI and accession numbers assigned to them by NCBI are KC741444, KC831587, KC831588, KC831589, KC831590, KC831591, and KC831592.

Antimicrobial assay

The *in vitro* antimicrobial activities of isolated extracts were tested on a group of significant Gram positive and Gram-negative bacteria (Table 1) and fungal strains (Table 2)

IIIM8 and IIIM2 was the most active of the all the isolated extracts against the bacterial pathogens. However the activity of IIIM8 and IIIM2 was limited to Gram-positive bacteria only as its MIC was

>256± µg/ml against *Escherichia coli* ATCC 25292 and *Pseudomonas aeruginosa* ATCC 27853 (Gram-negative pathogens used in this study). Whereas seven extracts IIIM3, IIIM4, IIIM5, IIIM6, IIIM7, IIIM1 and IIIM8 exhibited moderate Gram-positive antibacterial activity (MIC ≈ 32-128± µg/ml). Five extract IIIM9, IIIM10, IIIM11, IIIM12 and IIIM13 on the other hand were inactive against bacterial pathogens up to the tested concentration of 256± µg/ml. All the isolated extracts were completely devoid of antifungal activity up to the highest tested concentration of 256± µg/ml.

Table 1: Antimicrobial activity of the isolated extracts against bacterial pathogens

S. No	Tested Extracts	MIC µg/ml				
		<i>S. aureus</i> ATCC 29213	MRSA ATCC 15187	VRE	<i>E. coli</i> ATCC 25292	<i>P. aeruginosa</i> ATCC 27853
1	IIIM2	1	0.5	1	>256	>256
2	IIIM3	64	128	128	>256	>256
3	IIIM10	>256	>256	>256	>256	>256
4	IIIM4	64	64	64	>256	>256
5	IIIM5	32	32	64	>256	>256
6	IIIM6	32	32	32	>256	>256
7	IIIM11	>256	>256	>256	>256	>256
8	IIIM7	128	128	128	>256	>256
9	IIIM12	>256	>256	>256	>256	>256
10	IIIM13	>256	>256	>256	>256	>256
11	IIIM1	64	64	64	>256	>256
12	IIIM8	0.5	0.5	1	>256	>256
13	IIIM14	>256	>256	>256	>256	>256
14	IIIM9	>256	>256	>256	>256	>256
15	Cipro	0.25	16	64	0.03	0.03

Table 2: Antimicrobial activity of the isolated extracts against fungal pathogens

S. No	Tested Extracts	MIC µg/ml			
		<i>C. albicans</i> ATCC 90028	<i>C. albicans</i> V-0-191	<i>A. niger</i> 16404	<i>A. fumigates</i>
1	IIIM2	>256	>256	>256	>256
2	IIIM3	>256	>256	>256	>256
3	IIIM10	>256	>256	>256	>256
4	IIIM4	>256	>256	>256	>256
5	IIIM5	>256	>256	>256	>256
6	IIIM6	>256	>256	>256	>256
7	IIIM11	>256	>256	>256	>256
8	IIIM7	>256	>256	>256	>256
9	IIIM12	>256	>256	>256	>256
10	IIIM13	>256	>256	>256	>256
11	IIIM1	>256	>256	>256	>256
12	IIIM8	>256	>256	>256	>256
13	IIIM14	>256	>256	>256	>256
14	IIIM9	>256	>256	>256	>256
15	Ampho- B	0.5	0.5	1	1

DISCUSSION

In the present study we evaluated the antimicrobial potential of the endophytic strains both by agar diffusion assay and MIC determination assay against a series of microbial pathogens and eight isolates showed good activity against gram positive bacteria with two isolates showing promising activity with MIC in the range of 0.5 – 1($\mu\text{g/mL}$) in diameter .

To the best of our knowledge, for the first time we reported *in vitro* antimicrobial activities of endophytic fungal isolates of high value medicinal plants of Kashmir Valley, India. The reported fungal endophytes could be used as an appropriate source to produce antibacterial. Based on the study carried out it can be concluded that the two elite endophytic fungal strains, IIM2 and IIM8, have the potential to act as a source of new antibacterial compounds against pathogenic microorganisms (Gram positive bacteria and drug resistant bacteria). Roots of *R. emodi* are reported to have antibacterial and antifungal activities [13-15]. This strongly supports our results that the endophytes from these could be appropriate source of antimicrobial secondary metabolites and it is also evident from the study that fungal endophytes are a rich source of bioactive compounds [16-20]. Likewise, our study shows that fungal endophytes isolated from these plants, especially Kashmir valley India, also will be a rich array of antimicrobial compounds and warrant more intensive study in our search for new chemical structures. These reports and our results strongly support the view that the endophytic fungi of traditional medicinal plants are promising sources of natural antimicrobial compounds [21-23].

CONCLUSION

In an attempt made in the present study two endophytic fungi strains are expected to be a potential source for new natural bioactive molecules isolated from the high value medicinal plants of the Kashmir valley. These two endophytic fungal strains could be an alternative to the secondary metabolite production from medicinal plants and could a good reason to save the natural plant wealth of valley. These endophytic fungal strains could be a source of large scale production of antimicrobial compounds provided using fermentation studies.

Declaration of Interest

We the authors of the manuscript solely declare that the work done is original piece of work and has not been published anywhere or has not been submitted anywhere for publication. All the work has been properly cited as per literature. All the authors declare that there is no conflict of interests regarding the publication of this article. There are no issues regarding the authorship, financial and institutional affiliations. We authorize the corresponding author for all the matters regarding paper publishing should be communicating to him.

Acknowledgement

First author is thankful to CSIR, New Delhi, India for providing the financial assistance during the tenure of the work. We are also thankful to Dr. S. N. Sharma, Plant Systematic and Taxonomy, IIM, Jammu for identification and authentication of the medicinal plants.

REFERENCES

1. Fisher PJ, Andoson AE, Petrini O. Fungal endophytes in *Ulexeuropaeus* and *Ulexgalli*. *Trans Br. Mycol. Soc* 1986;86:153-56.
2. Bacon CW. Isolation,culture and maintenance of endophytic fungi of grasses In: Isolation of biotechnological organisms from nature. ed. by D.P. Labeleda, 1990, McGraw-Hill, New York, USA.
3. Petrini O. Fungal endophyte of tree leaves. In: Andrews J, Hirano SS (eds) *Microbial ecology of leaves*. Springer, New York, 1992; 179-197.
4. Strobel GA, David ML. Endophytic microbes embody pharmaceutical potential. *ASM News* 1998;64:263-68.
5. Strobel GA, Daisy B. Bioprospecting for Microbial Endophytes and Their Natural Products. *Microbiology and Molecular Biology Reviews* 2003;67:491-502.
6. Tan RX, Zou WX. Endophytes: a rich source of functional metabolites. *Natural Product Reports* 2001;18:448-59.
7. Stroble G. Endophytes as source of bioactive products. *Microbes and infection*. 2003;5:534-44.
8. Strobel GA, Dirksie E, Sears J, Markworth C. Volatile antimicrobials from a novel endophytic fungus. *Microbiol* 2001;147:2943-50.
9. Azevedo JL, Maccheroni JRW, Pereira JO, Araújo WL. Endophytic microorganisms: a review on insect control and recent advances on tropical plants. *Eletronic Journal of Biotechnology* 2000;3:40-65.
10. Arnold AE, Maynard Z, Gilbert GS, Coley PD, Kursar TA. Are tropical fungal endophytes hyper-diverse? *Ecology Letters* 2000;3(4):267-74.
11. Vainio EJ, Korhonen K, Hantula J. Genetic variation in *Phlebiopsis gigantea* as detected with random amplified microsatellite (RAMS) markers. *Mycol Res* 1998;2:187-92.
12. Wayne PA. Clinical and Laboratory Standards Institute. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. 7th ed. Approved standard. 2006;M7-A7.
13. Agarwal JS, Rastogi RP, Srivastava OP. In vitro toxicity of constituents of *Rumexmaritimus*Linn. to ringworm fungi. *Curr Sci*, 1976;45:619-20.
14. Cyong JC, Matsumoto T, Arakawa K, Kiyohara H, Yamada H, Otsuka Y. Antibacteriosides *Fragilus* substance from rhubarb. *Journal of Ethnopharmacology*. 1987;19:279-83.
15. Harvey HE, Waring JM. Antifungal and other compounds isolated from the roots of New Zealand flax plants (The genus *Phormium*). *J Nat Prod*, 1987;50, 767-76.
16. Bills GF, Polishook JD. Recovery of endophytic fungi from *Chamaecyparisthyoides*. *Sydowia* 1992;44:1-12.
17. Rodrigues KF, Petrini O. Biodiversity of endophytic fungi in tropical regions. In: (K.D. Hyde, ed.). *Biodiversity of tropical fungi*. Hong Kong University Press, Hong Kong. 1997;57-69.
18. Schulz B, Boyle C, Draeger S, Römmert A, Krohn K.. Endophytic fungi: a source of novel biologically active secondary metabolites. *Mycological Research* 2002;106: 996-1004.
19. Stroble G. Endophytes as source of bioactive products. *Microbes and infection*. 2003; 5: 534-44.
20. Stadler M, Hellwig V. Chemotaxonomy of the Xylariaceae and remarkable bioactive compounds from Xylariales and their associated asexual stages. *Recent Res. Dev. Phytochem*. 2005;9: 41-93.
21. Stierle A, Strobel G, Stierle D. Taxol and taxane production by *Taxomyces*, an endophytic fungus of pacific yew (*Taxus brevifolia*). *Science* 1993;260:214-16.
22. Strobel GA, Torczynski R, Bollon A. *Acremonium* sp. a leucinostainA producing endophyte of European yew (*Taxus baccata*). *Plant Science* 1997;128:97-108.
23. Strobel GA, Daisy B. Bioprospecting for microbial endophytes and their natural products. *Microbiology and Molecular Biology Reviews* 2003;67:491-502.

HOW TO CITE THIS ARTICLE

Dar RA, Saba I, Shah Nawaz M, Qazi PH, Khan IA. Antimicrobial potential of fungal endophytes from selected high value medicinal plants of the Kashmir valley – India. *J Phytopharmacol* 2017;6(5):307-310.