The Journal of Phytopharmacolog (Pharmacognosy and phytomedicine Research)

Research Article

ISSN 2320-480X JPHYTO 2020; 9(5): 367-370 September- October Received: 31-08-2020 Accepted: 11-10-2020 ©2020, All rights reserved doi: 10.31254/phyto.2020.9513

BN Satish

PhD Scholar and Professor, Sri Dharmasthala Manjunatheshwara College of Ayurveda, Kuthpady, Udupi, Karnataka, India

Mallya Suma V

Associate Professor, Sri Dharmasthala Manjunatheshwara College of Ayurveda, Kuthpady, Udupi, Karnataka, India

Vishwanatha

Research Officer, Sri Dharmasthala Manjunatheshwara College of Ayurveda, Kuthpady, Udupi, Karnataka, India

Correspondence: Dr. Mallya Suma V

Associate Professor, Sri Dharmasthala Manjunatheshwara College of Ayurveda, Kuthpady, Udupi, Karnataka, India Email: sumamallya@gmail.com

Screening for cytotoxic activity of *Habenaria* longicorniculata J graham tubers- an *in vitro* study

BN Satish, Mallya Suma V, Vishwanatha

ABSTRACT

About: *Habenaria longicorniculata* J. Graham are tuberous orchid, the tubers utilized by flok healers in cancer managemnet, as a rejuvenator. A study has been planned to evaluate *In-vitro* cytotoxicity of tuber extract against selected cell lines. **Materials and Methods:** *H. longicorniculata* J.Graham identified, uprooted during their flowering time. Tuber extract of this plant used for its *In-vitro* cytotoxicity against selected cell lines of Human Breast cancer (MCF 7), Human Liver carcinoma (HepG2), and Human cervix adenocarcinoma (HeLa) cells as per standard protocol. **Results:** Tuber Extract exhibited a CTC₅₀ value of >1000 on MCF 7, HepG2 and HeLa cell lines. The results from the MTT assay indicate that 72hr extract incubation with the combined extracts is toxic to the cells and the level of damage is concentration dependent.

Keywords: *Habenaria longicorniculata, In-vitro* study, Cytotoxic activity, Human Breast cancer (MCF 7), Human Liver carcinoma (HepG2), Human cervix adenocarcinoma (HeLa).

INTRODUCTION

Changed life style, pollution, stress related factors are prime cause of increased incidence of malignant disorders globally ^[1]. Secondary metabolites of few plant species have proved toxic on these cancer cells, simultaneously not affecting other normal cells². Many plants have been screened to test their efficacy in cancer since 1950 ^[3]. A great number of plant species have been investigated for cytotoxic, antitumour and anticancer activities which included both in-vitro and in-vivo models ^[4]. These preclinical studies have significant contribution in novel drug discovery in cancer treatment, incorporating a variety of cell lines and tumor types in the pre-screening, screening protocols of potential anticancer drugs ^[5]. As cancer was primarily considered a disease of uncontrolled cell division, by measuring the regression in tumor size, identification of a cytotoxic or an antiproliferative compound was considered as the main objective endpoint of efficacy of a compound in preclinical and clinical anticancer drug development for decades ^[6]. A potent anticancer drug must kill or weaken cancer cells without injuring normal cells ^[7]. Traditional/ verbal community has a vast knowledge on various anticancer drugs, but pharmacological action yet to be derived ^[8].

Habenaria longicorniculata J. Graham is an orchid, found at peninsular India belonging to family *Orchidaceae*, found commonly at hilly, slope areas among grasses ^[9]. Orchid usually gives white flowers in the month of August, during which underground tubers to be collected ^[10]. Traditional physicians use these tubers in the treatment of malignancy, as a rejuvenator also in many ailments ^[11]. Tubers said to be beneficial in wasting disease, fever, disorders of blood, haemorrhage and specifically used as overall rejuvenator ^[12]. The tubers are said to possess antioxidant property. Today whole world is looking for scientific evidences for traditional claims. Hence a study has been planned to evaluate *In-vitro* cytotoxicity of tuber extract against selected cell lines of Human Breast cancer (MCF 7), Human Liver carcinoma (HepG2), and Human cervix adenocarcinoma (HeLa) cells.

Cell culture technique an exclusive research in cellular and molecular biology, which provide exceptional study design in the field of normal physiology and biochemistry of cells (e.g., metabolic studies, aging)¹³. This is also used in new herb screening, drug preparation (e.g., vaccines, therapeutic proteins). Uniqueness in consistency and reproducibility of results using batch of clonal cells, make it popular among invitro study¹⁴. Thus based on above all mentioned factor a study has been designed to evaluate In-vitro cytotoxic activity of *Habenaria longicorniculata* J.Graham tubers on few malignant cell lines.

MATERIALS AND METHODS

Tubers of *Habenaria longicorniculata* J. Graham were collected during their flowering season, from their natural habitat. Plant sample cleaned using water, authenticated, roots detached from plant and shade dried.

Tuber extract taken for its *In-vitro* cytotoxicity against selected cell lines of Human Breast cancer (MCF 7), Human Liver carcinoma (HepG2), and Human cervix adenocarcinoma (HeLa) cells ^[15].

METHODOLOGY

Test substance was prepared at concentrations ranging from 1000 μ g/ml to7.8 μ g/ml to determine the percentage growth inhibition of the test substance on MCF 7, HepG2 and HeLa cell lines ^[16].

Preparation of test solution

For cytotoxicity studies, 10mg of test drug was separately weighed and volume was made up with DMEM-HG/MEM supplemented with 2% inactivated FBS to obtain a stock solution of 10% w/v concentration and sterilized by 0.22 μ syringe filtration. Serial two-fold dilution was arranged from this for carrying out cytotoxicity studies ^[17].

Cell lines and Culture medium

Human breast cancer (MCF 7), Human Liver carcinoma (HepG2), and Human cervix adenocarcinoma (HeLa) cell lines were obtained from National Centre for Cell Sciences (NCCS), Pune, India. Stock cells were cultured in DMEM-HG/MEM supplemented with 10% inactivated Fetal Bovine Serum (FBS), penicillin (100 IU/ml), streptomycin (100 µg/ml) and amphotericin B (5 µg/ml) in a humidified atmosphere of 5% CO₂ at 37°C until confluent. The cells were dissociated with TPVG solution (0.2% Trypsin, 0.02% EDTA, 0.05% glucose in PBS). The stock cultures were grown in 25 cm² culture flasks and all experiments were carried out in 96 well microtitre plates (Tarsons India Pvt. Ltd., Kolkata, India) ^[18].

Cytotoxicity Study

The monolayer cell culture was trypsinized and the cell count was adjusted to 100,000 cells/ml using DMEM-HG/MEM containing 10% FBS. To each well of the 96 well microtitre plate, 0.1 ml of the diluted cell suspension was added. After 24 hours, when a partial monolayer was formed, the supernatant was skimmed off, the monolayer washed once with medium and 100 µl of different test concentrations of test drugs were added on to the partial monolayer in microtitre plates. The plates were then incubated at 37°C for 72 hr in 5% CO2 atmosphere, and microscopic examination was carried out and observations were noted every 24hr interval. After 72 hr, the drug solutions in the wells were thrown off and 50 µl of MTT in PBS was added to each well. The plates were gently shaken and incubated for 3 hr at 37°C in 5% CO₂ atmosphere. The supernatant was removed and 100 µl of propanol was added and the plates were gently shaken to solubilize the formed formazan. The absorbance was measured using a microplate reader at a wavelength of 540 nm. The percentage growth inhibition was calculated using the Standard formula and concentration of test drug needed to inhibit cell growth by 50% (CTC₅₀) values was generated from the dose-response curves for each cell line [19].

RESULTS

Tuber extract of *Habenaria longicorniculata* J Graham showed results for MCF 7 Human breast carcinoma cells, HepG2 Human Liver carcinoma cells, HeLa Human Cervical adenocarcinoma cells as follows. The percentage growth inhibition was calculated using the Standard formula and concentration of test drug needed to inhibit cell growth by 50% (CTC₅₀) values was generated from the dose-response curves for each cell line. 1. Cytotoxic properties against MCF 7 cell line:

At the test concentration of 1000, 500, 250, 125, 62.5, 31.25, 15.5 and 7.8, the Percentage of Cytotoxicity inhibition was 37.34, 32.87, 26.40, 18.75, 13.19, 11.70, 10.28 and 1.88 respectively. The percentage growth inhibition was calculated using the Standard formula and concentration of test drug needed to inhibit cell growth by 50% (CTC_{50}) values was generated from the dose-response curves for each cell line. (Table 1)

Table 1: Cytotoxic properties of *Habenaria longicorniculata* J Graham

 against MCF 7 cell line

| Sl. | Test | Test Conc. | % | CTC ₅₀ |
|-----|---------------|------------|---------------|-------------------|
| No. | Compound | (µg/ml) | Cytotoxicity | (µg/ml) |
| 1. | Tuber Extract | 1000 | 37.34±0.38 | |
| | | 500 | 32.87±0.18 | |
| | | 250 | 26.40±0.24 | |
| | | 125 | 18.75±0.22 | >1000 |
| | | 62.5 | 13.19±0.29 | |
| | | 31.25 | 11.70±0.30 | |
| | | 15.5 | 10.28±0.30 | |
| | | 7.8 | 1.88 ± 0.11 | |

2. Cytotoxic properties against HepG2 cell line

At the test concentration of 1000, 500, 250, 125, 62.5, 31.25, 15.5 and 7.8, the Percentage of Cytotoxicity inhibition was 33.07, 24.75, 17.30, 13.59, 12.51, 07.45, 02.15 and 0.82 respectively. (Table 2)

Table 2: Cytotoxic properties of *Habenaria longicorniculata* J Graham

 against HepG2 cell line

| Sl. No. | Name of Test Compound | Test Conc. (µg/ml) | % Cytotoxicity | СТС ₅₀ (µg/ml) |
|------------|--------------------------|-----------------------|-------------------|------------------------------|
| 1. | Tuber Extract | 1000 | 33.07±0.31 | |
| | | 500 | 24.75±0.28 | |
| | | 250 | 17.30±0.20 | |
| | | 125 | 13.59±0.36 | >1000 |
| | | 62.5 | 12.51±0.52 | |
| | | 31.25 | 7.45±0.31 | |
| | | 15.5 | 2.15±0.27 | |
| | | 7.8 | 0.82 ± 0.46 | |

3. Cytotoxic properties against HeLa cell line

At the test concentration of 1000, 500, 250, 125, 62.5, 31.25, 15.5 and 7.8, the Percentage of Cytotoxicity inhibition was 45.81, 41.74, 40.20, 38.64, 36.48, 35.09, 24.96 and 8.04 respectively. (Table 3)

 Table 3: Cytotoxic properties of Habenaria longicorniculata J Graham

 against HeLa cell line

| Sl. No. | Name of Test Compound | Test Conc. (µg/ml) | % Cytotoxicity | CTC ₅₀ (µg/ml) |
|------------|--------------------------|-----------------------|-------------------|------------------------------|
| 1. | Tuber Extract | 1000 | 45.81±0.68 | |
| | | 500 | 41.74±0.38 | |
| | | 250 | 40.20±0.16 | |
| | | 125 | 38.64±0.43 | >1000 |
| | | 62.5 | 36.48±0.32 | |
| | | 31.25 | 35.09±0.63 | |
| | | 15.5 | 24.96±0.78 | |
| | | 7.8 | 8.04 ± 0.47 | |



Figure 1: Graph of cytotoxic effect of test substance on MCF 7.



Figure 2: Graph of cytotoxic effect of test substance on HepG2



Figure 3: Graph of cytotoxic effect of test substance on HeLa cell line.

DISCUSSION

Herbs have been used since centuries as therapeutics to cure illness as well as to build up immunity. Plants have a major role in maintaining health of this earth as well all living beings. *In vitro* cytotoxicity screening models afford significant introductory data to help select plant extracts with probable anticancerous activity²⁰. Although *Habenaria longicorniculata* J.Graham tubers have valuable pharmacological effects, the comprehensive awareness about its cytotoxic activity has been lacking.

In-vitro cytotoxicity of Tubers extract of *Habenaria longicorniculata* J Graham was tested against MCF 7, HepG2 and HeLa cell line. Test substance was taken at concentrations ranging from 1000 μ g/ml to 7.8 μ g/ml to determine the percentage growth inhibition of the test

substance on MCF 7, HepG2 and HeLa cell lines. The test substance extract exhibited a CTC_{50} value of >1000 on MCF 7, HepG2 and HeLa cell lines.

The MTT assay is a delicate, measurable and consistent colorimetric assay that measure cell capability²¹. This is based on the capacity of the cellular mitochondrial dehydrogenase enzyme in living cells to reduce the yellow water-soluble substrate 3-(4,5-dimethylthiazol-2yl)-2,5-diphenyl tetrazolium bromide (MTT) into a dark blue/purple formazan product which is insoluble in water. The amount of formazan produced is directly proportional to the cell number in a range of cells lines. According to the American National Cancer Institute (NCI), the criteria of cytotoxicity for crude extracts is an $CC_{50} < 30 \mu g/mL$ after an exposure time of 72 h in a preliminary assay ^[22]. Tuber Extract of *Habenaria longicorniculata* J Graham met these criteria with CC_{50} value less than 30 $\mu g/mL$ with the crude extract being the most cytotoxic.

Test drug extract (HL) showed results for MCF 7 Human breast carcinoma cells at the test concentration of 1000 μ g/ml - 37.34% and at 500 μ g/ml - 32.87% of cytotoxicity inhibition was seen respectively. Whereas for HepG2 Human Liver carcinoma cells at the test concentration of 1000 μ g/ml - 33.07% and at 500 μ g/ml - 24.75% of cytotoxicity inhibition was observed. For HeLa Human Cervical adenocarcinoma cells at the test concentration of 1000 μ g/ml - 45.81% and at 500 μ g/ml - 41.74% of cytotoxicity inhibition was found by test drug (HL).

The results from the MTT assay indicate that 72hr extract incubation with the combined extracts is toxic to the cells and the level of damage is concentration dependent. Unwanted reactions can give rise to high background absorbance values especially in herbal extracts when antioxidants in the extracts directly react with MTT or formazan. This occurrence was minimal in these experiments because the blanks (media without cells) did not give background reactions. Test drug extract has shown considerable cytotoxic activity on malignant cells, thus proving its anticancer property.

CONCLUSION

Tuber Extract of *Habenaria longicorniculata* J Graham was tested for *In vitro* cytotoxicity studies against MCF 7 (Human breast cancer), Human Liver carcinoma (HepG2), and Human cervix adenocarcinoma (HeLa) cells by MTT assay exposing the cells to different concentrations of test substance. Tuber Extract (HL) exhibited a CTC_{50} value of >1000 on MCF 7, HepG2 and HeLa cell lines. The results from the MTT assay indicate that 72hr extract incubation with the combined extracts is toxic to the cells and the level of damage is concentration dependent. Thus, a promising anticancer drug, to confirm with further experimental models.

Conflict of interest

The authors declare no conflict of interests

Funding

No funding was received for this study.

REFERENCES

 Ali Imran, Wani A Waseem, Salem Kishwar. Cancer Scenario in India with Future perspectives; Cancer Therapy. 2011; 8(8):56-70

- Supriya Korrapati, Pallavi Korra, Srinivasababu Puttugunta; Natural and Herbal Remedies for Cancer Treatment; Inventi Impact; Planta Activa, 2016; (4):140-7
- Evans WC., Trease and Evan's Pharmacognosy, 15th ed. London: WB Saunders Ltd: 2002, 472-85
- Kinghorn A. Plant Secondary Metabolites as Potential Anticancer Agents and Cancer Chemopreventives. Molecules. 2000; 5(3):285-8.
- Sachin Kumar, Sakshi Bajaj, Ramesh Bodla; Preclinical screening methods in cancer; Indian Journal of Pharmacology; September 2016; 48(5):481-486
- Monks *et al.* Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines J Natl. Cancer Inst. 1991; 83:757-766
- WHO, Diet, Nutrition and the Prevention of Chronic Diseases, [World Health Organization, Technical Report Series, 916, WHO, Geneva, 2003. Phillips HJ and Terryberry JE. Counting actively metabolizing tissue cultured cells. Cell Research 1957; 13:341-347.
- 8. Mukharjee Pulok K, Quality Control of Herbal Drugs; New Delhi: Business Horizons, 2002, 68
- Satish Bn, Mallya Suma V, Prabhu Suchitra. Authentication parameters of *Habenaria longicorniculata* J. Graham; medicinal orchid used in Ayurveda; Arya vaidyan, 2018; XXXI(3):31-40.
- BN Satish, Mallya Suma V, *In vitro* antioxidant activity assay of *Habenaria longicorniculata* J. Graham. Wild medicinal tubers; Journal of Drug Delivery and Therapeutics; 2019; 9(3):342-344.
- 11. Kirtikar KR, Basu BD. Indian Mediicnal Plants, Volume III, Dehradun; International Book Distributors; 2008, 2399-415
- 12. BN Satish, Mallya Suma V., Differentiation and authentication of medicinal orchids of *Habenaria* species using RAPD- a molecular profile; WJPR, 2018; 7(17):886-92.
- Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D et al. Evaluation of Colorimetric Protein and Biomass Stains for Assaying Drug Effects upon Human Tumor Cell Lines. Proceedings of the American Association for Cancer Research 1989; 30:612.
- Masters RW. Animal cell culture, Trypan Blue Assay sop. 3rd ed. 2000, 1-3.
- 15. Li XY, Wang X. The role of human cervical cancer oncogene in cancer progression. *Int J Clin Exp Med.* 2015; 8(6):8363-8368.
- Cogswell PC, Guttridge DC, Funkhouser WK, Baldwin AS Jr. Selective activation of NF kappa B subunits in human breast cancer: potential roles for NF-kappa B2/p52 and for Bcl-3. *Oncogene*. 2000; 19(9):1123-1131.
- Choi HJ, Lee JM, Kim H, *et al.* Bcl3-dependent stabilization of CtBP1 is crucial for the inhibition of apoptosis and tumor progression in breast cancer. *Biochem Biophys Res Commun.* 2010; 400(3):396-402.
- Ekalaksananan T, Sookmai W, Fangkham S, *et al.* Activity of androgra¬pholide and its derivatives on HPV16 pseudovirus infection and viral oncogene expression in cervical carcinoma cells. Nutr Cancer. 2015;67(4):687-696.
- Maldonado V, Espinosa M, Pruefer F, *et al.* Gene regulation by BCL3 in a cervical cancer cell line. *Folia Biol (Praha).* 2010; 56(4):183-193.
- Qureshi R, Arora H, Rizvi MA. EMT in cervical cancer: its role in tumour progression and response to therapy. *Cancer Lett.* 2015; 356(2 Pt B):321-331.
- Mark Shoemaker, Isan Cohen, Micheal Campbell; Reduction of MTT by aqueous herbal extracts in the absence of cell, Journal of Ethnopharmacology, Vol 93(2-3) August 2004, 381-84

22. Farhondeh Nemati *et al.*, Cytotoxic properties of some Medicinal plant extracts from Mazandaran, Iran; Iran Red Cresent Medical Journal. 2013; 15(11):871.

HOW TO CITE THIS ARTICLE

Satish BN, Mallya SV, Vishwanatha. Screening for cytotoxic activity of *habenaria longicorniculata* J graham tubers-an *in vitro* study. J Phytopharmacol 2020; 9(5):367-370.