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## Research Article

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## Network Pharmacology study on the mechanism of MKA Polyherbal Formulation in combating Respiratory Diseases

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### ABSTRACT

The Multi-targeted action of Polyherbal formulation is responsible for enhanced therapeutic efficacy in combating various diseases. But, understanding the mode of action of herbal medicine remains a challenge because of its complex metabolomics. Network pharmacology-based approach enables to explore the mechanism of action of polyherbal formulation in biological system. In present investigation, we have explored the molecular mechanism of action of the Polyherbal formulation MKA comprising of three botanicals *Mimusops elengi* L., *Kedrostis foetidissima* (Jacq.) Cogn. and *Artemisia vulgaris* L. in treating respiratory diseases by network pharmacology-based approach. The protein targets were mined from Binding database for the bioactive present in MKA. The disease associated targets were identified using Open target Platform. Based on ligand-target interactions, it was interpreted that MKA could alleviate the symptoms of respiratory disease by multiple mechanisms like EGFR inhibition by Quercetin and Quercetin-3-O-rhamnoside, KDR inhibition by Quercetin, STAT-3 inhibition by  $\beta$ -sitosterol-  $\beta$ -D-glucoside, TRPV1 inhibition by phytol acetate, etc. The Protein-protein interaction (PPI) network was constructed using STRING database. KEGG pathway based functional enrichment was also predicted for the PPI network. It was found that multiple ligand-target interactions and protein-protein interactions is responsible for pharmacological activity of MKA in respiratory diseases.

**Keywords:** Polyherbal formulation, Respiratory, Ligand, Protein-protein interactions (PPIs), Inhibition.

### INTRODUCTION

Respiratory diseases are the most common cause of illness and death all over the world. Among the respiratory diseases, asthma, chronic obstructive pulmonary disease (COPD) and acute respiratory distress syndrome (ARDS) are related to high mortality rate [1,2,3]. The main causative agents of these infectious diseases are virus and bacteria. SARS-CoV infection, mainly targets the respiratory tract leading to chronic inflammation and arrests the lung function [4,5]. In most of the cases, a single compound extracted and purified from polyherbal formulation or traditional medicine may have lower activity compared to the formulation, and in certain cases, the single compound extracted may be more toxic. This confirms that, there is a difference in activity of the compound individually and as such in the formulation. So, synergistic action of compounds in polyherbal formulation has to be considered for its therapeutic value [6,7].

Network pharmacology is a newly emerging area which deals with relationship between ligands, protein targets and its associated diseases in biological network of herbal formulations. It clearly demonstrates the synergistic action of multiple ligands on multiple targets in herbal formulations [8]. The MKA Polyherbal formulation comprising of three selected plants *Mimusops elengi*, *Kedrostis foetidissima* (Jacq.) Cogn. and *Artemisia vulgaris* L. were reported to possess anti-inflammatory potential in LPS induced RAW 264.7 macrophages. The MKA treatment decreased the release of Reactive Oxygen Species (ROS) and nitrite level in inflammatory macrophages [9]. Also, the MKA was explored to possess therapeutic effect on LPS treated macrophages by reducing the rate of lipid peroxidation and restoring the mitochondrial membrane potential and cell membrane integrity [10].

In present investigation, network pharmacological method was employed to understand the molecular mechanisms underlying the action of three botanicals *Mimusops elengi*, *Kedrostis foetidissima* (Jacq.) Cogn. and *Artemisia vulgaris* L. in MKA herbal formulation in combating respiratory diseases. The potential protein targets of the bioactives present in polyherbal formulation MKA were identified from binding database. The respiratory disease associated protein targets were identified from Open target platform. The protein-protein interactions (PPIs) were interpreted by constructing the protein network map using STRING database.

## MATERIALS AND METHODS

### Mining of phytochemicals and Protein targets

The bioactive compounds present in each plant constituent of Polyherbal formulation MKA were mined from scientific literatures, journals and biological databases. In total, five research reports were used as references to study the phytochemicals in *Mimusops elengi* L., *Kedrostis foetidissima* (Jacq.) Cogn. and *Artemisia vulgaris* L. [11-15].

The PUBCHEM Compound ID and canonical SMILES for all the chemical compounds were retrieved from PUBCHEM database (<https://pubchem.ncbi.nlm.nih.gov/>) [16]. The chemical compounds without PUBCHEM Compound ID were excluded. The Potential protein targets were identified from binding database (<https://www.bindingdb.org/bind/index.jsp>) using CANONICAL SMILES as query input. The targets having at least 70% ligand template similarity were selected for further studies [17]. The ligand target interaction was also predicted from Binding database based on the assay description.

### Respiratory disease associated target identification

The target proteins associated with respiratory diseases were predicted from open targets (<https://www.targetvalidation.org/>), which is a platform for therapeutic target identification and validation [18]. The lists of proteins identified from binding database were given as input using "Batch search" option. The Open Target output, gives the details

of the relevant disease associated proteins. The respiratory disease associated protein targets were screened from the Open Target output. The affinity of the ligand present in MKA with respiratory disease associated protein targets was analyzed.

### Construction of PPI network

The metabolic interactions of the protein targets associated with respiratory diseases were studied using STRING database. The list of proteins identified from Open target Platform associated with respiratory disease, was given as input in STRING database. Multiple protein search was conducted to predict the protein-protein interactions (PPIs) network [19].

## RESULTS AND DISCUSSION

### Phytoconstituents present in MKA Polyherbal formulation

The phytoconstituents present in individual plants *Mimusops elengi* L., *Kedrostis foetidissima* (Jacq.) Cogn. and *Artemisia vulgaris* L. of MKA were identified by mining of literatures and databases. The chemical compounds with their PUBCHEM CID and CANONICAL SMILES were illustrated in Table 1. It is inferred that, MKA is a rich source of phytochemicals which is responsible for its therapeutic effect. For present investigation, 13 bioactives from *Mimusops elengi* L., 14 bioactives from *Kedrostis foetidissima* (Jacq.) Cogn. and 13 phytochemicals from *Artemisia vulgaris* L. were used for prediction of multi-target pharmacological mechanism of action of MKA Polyherbal formulation in treating respiratory diseases.

**Table 1:** Bioactives present in MKA Polyherbal formulation

BOTANICALS	LIGANDS	PUBCHEM ID	CANONICAL SMILES
<i>Mimusops elengi</i> L.	Quercetin	5280343	<chem>C1=CC(=C(C=C1)C2=C(C(=O)C3=C(C=C(C=C3O2)O)O)O)O</chem>
	Quercitol	441437	<chem>C1C(C(C(C1O)O)O)O</chem>
	$\beta$ -sitosterol- $\beta$ -D-glucoside	71628	<chem>CCC(CCC(C)C1CCC2C1(CCC3C2CC=C4C3(CCC(C4)OC5C(C(C(C(O5)CO)O)O)C)C)C)C</chem>
	Ursolic acid	64945	<chem>CC1CCC2(CCC3(C=CCC4C3(CCC5C4(CCC(C5(C)C)O)C)C)C2C1C)C(=O)O</chem>
	Stearic acid 3-(octadecyloxy) propyl ester	551406	<chem>CCCCCCCCCCCCCCCCCCOCCCC(=O)CCCCCCCCCCCCCCCCCC</chem>
	Hexadecanoic acid, methyl ester	8181	<chem>CCCCCCCCCCCCCCCCCC(=O)OC</chem>
	10-Octadecenoic acid, methyl ester	25642	<chem>CCCCCCCC=CCCCCCCCC(=O)OC</chem>
	Spinasterol	5281331	<chem>CCC(C=CC(C)C1CCC2C1(CCC3C2=CCC4C3(CCC(C4)O)C)C)C(C)C</chem>
	Hentriacontane	12410	<chem>CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC</chem>
	$\beta$ -carotene	5280489	<chem>CC1=C(C(CCC1)(C)C)C=CC(=CC=CC(=CC=CC=C(C)C)C=CC=C(C)C=CC2=C(CCCC2(C)C)C)C</chem>
	Stearic acid 3-(octadecyloxy) propyl ester	551406	<chem>CCCCCCCCCCCCCCCCCCOCCCC(=O)CCCCCCCCCCCCCCCCCC</chem>
	Squalene	638072	<chem>CC(=CCCC(=CCCC(=CCCC=C(C)CCC=C(C)CCC=C(C)C)C)C)C</chem>
	$\beta$ -sitosterol	222284	<chem>CCC(CCC(C)C1CCC2C1(CCC3C2CC=C4C3(CCC(C4)O)C)C)C(C)C</chem>
<i>Kedrostis foetidissima</i> (Jacq.) Cogn.	Quercetin-3-O-rhamnoside	5280459	<chem>CC1C(C(C(C(O1)OC2=C(C(OC3=CC(=CC(=C3)O)O)O)C4=CC(=C(C=C4)O)O)O)O)O</chem>
	7,10-Hexadecadienoic acid, methylester (CAS)	548878	<chem>CCCCCC=CCC=CCCCCCC(=O)OC</chem>
	Eicosanoic acid, methyl ester(CAS)	14259	<chem>CCCCCCCCCCCCCCCCCCCC(=O)OC</chem>
	Phytol Acetate	6428538	<chem>CC(C)CCCC(C)CCCC(C)CCCC(=CCOC(=O)C)C</chem>
	Curcubitacin	181183	<chem>CC(=O)OC(C)C(C)CCC(=O)C(C)C1C(CCC2(C1(CC(=O)C3(C2CC=C4C3CC(C(C4(C)C)O)O)C)C)O)O</chem>

	1H-Indole, 1-methyl-(CAS)	11781	CN1C=CC2=CC=CC=C21
	Rutin	5280805	CC1C(C(C(C(O1)OCC2C(C(C(C(O2)OC3=C(OC4=CC(=CC(=C4C3=O)O)O)C5=CC(=C(C=C5)O)O)O)O)O)O)O
	Docosanoic acid	8215	CCCCCCCCCCCCCCCCCCCC(=O)O
	9-Octadecenoic acid, methyl ester	5280590	CCCCCCCC=CCCCCCCC(=O)OC
	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]-(CAS)	5366244	CC(C)CCCC(C)CCCC(C)CCCC(=CCO)C
	1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester	6782	CC(C)COC(=O)C1=CC=CC=C1C(=O)OCC(C)C
	1H-1,2,4-Triazole-3,5-dicarbaldehyde	638657	C(=O)C1=NC(=NN1)C=O
	m-carboxyphenyl alanine	57370423	CC(C(=O)O)N(C1=CC=C(C=C1)C(=O)O)C(=O)OC(C)C
	Citrulline	833	C(CC(C(=O)O)N)CNC(=O)N
<i>Artemisia vulgaris L.</i>	chavicol	68148	C=CCC1=CC=C(C=C1)O
	borneol	64685	CC1(C2CCC1(C(C2)O)C)C
	Osopulegol	24585	CC1CCC(C(C1)O)C(=C)C
	myrtenol	10582	CC1(C2CC=C(C1C2)CO)C
	Camphor	2537	CC1(C2CCC1(C(=O)C2)C)C
	p-Cymene	7463	CC1=CC=C(C=C1)C(C)C
	1,8-Cineole	2758	CC1(C2CCC(O1)(CC2)C)C
	α-Pinene	440968	CC1=CCC2CC1C2(C)C
	β-Caryophyllene	5281515	CC1=CCCC(=C)C2CC(C2CC1)(C)C
	(Z)-β-Farnesene	5317319	CC(=CCC/C(=C\CCC(=C)C=C)/C)C
	Trans-2-Hexenal	5281168	CCCC=CC=O
	β-Pinene	440967	CC1(C2CCC(=C)C1C2)C
	D-Limonene	440917	CC1=CCC(CC1)C(=C)C

**Table 2:** Potential protein targets of Phytoconstituents present in MKA

PHF	Botanicals	Protein targets
MKA	<i>Mimusops elengi L.</i>	SCD, FABP1, CACNA1S, FABP3, FABP2, SLC22A20P, FLT3, GLO1, PRKCH, F7, HSD11B2, OSBP, CYP17A1, CYP19A1, CYP51A1, AAAS, ESR2, NR1H2, RORC, PTPN1, SREBF2, SHBG, PTPN2, HSD11B1, HMGCR, AR, ALOX15B, CD81, CDC25B, PTPRC, ELANE, PPARA, PTGES2, HINT1, CCT7, SLC01B1, SLC01B3, SRC, CA2, CA4, CA6, CA1, NR6A1, RXRG, RARA, ADORA1, RXRA, ADORA2A, ADORA3, ADORA2B, VDR, STAT3, POLL, DUSP3, PTPN7, GLS, MRPL57, IL2, BCL2L1, UBE2N, ABCC4, AKR1B10, TOP2A, TOPBP1, CRHBP, CYP3A5, ALOX15, MAOA, ALOX5, AHR, ABCG2, AURKB, CA12, CA7, CISD1, CDK1, CDK5, CDK6, PTGS1, CYP1A1, CYP1B1, ELAVL3, EGFR, HSD17B2, MET, IGF1R, MMP12, NOX4, PI4KA, PIK3CG, PIM1, TTR, TYR, LCK, AXL, KDR, XDH, PRKCA, FNTA, FABP4, FABP5, SLC22A11, TLR2, DNMT1, GPR84, FFAR1, FAAH, FFAR4, NR1I2, MAOB
	<i>Kedrostis foetidissima (Jacq.) Cogn.</i>	ITGAL, ARHGAP35, PRKD1, F10, SCD, F7, ATG4B, MAOB, FABP1, FABP3, FABP2, CA4, CA6, CA1, CA2, KIF23, NQO2, CDK4, DDC, CDK2, AAAS, ADK, HTR2A, HTR2C, HTR6, HSP90AA1, PTPN1, TRPV1 (VR1), FABP4, FABP5, AKR1B10, SLC22A11, TLR2, DUSP3, PTPN7, DNMT1, GPR84, FFAR1, FNTA, ALOX5, GSK3B, CDC25A, OXER1, PTGS1, TECRL, CYP19A1, PLA2G15, FAAH, LTB4R, TOPBP1, SOAT1, FFAR4, PPARA, NR1I2, SLC6A4, CTSB, CTSK, DDAH1, NOS3, NOS1, NOS2, CYP3A5, ADORA3, ADORA2A, CA12, CA7, PDE6C, CHRM5, EGFR, IL2, ALPI, NOX4, AMY2A, RPS6KA3, TNF, HSD11B1, AKR1C1, CTRC, CYP17A1, ESR2, NR3C1, NR3C2, PLA2G2A, PGR, PTGES2, PRKCA, PRKCD, CDC42BPA, CDC42BPB, SHBG, PRSS1
	<i>Artemisia vulgaris L.</i>	NR1H3, NR1H2, PRKCB, PRKCA, G6PD, F12, SOAT1, MAOA, MAOB, TRPA1, CNR2, CNR1, FAAH, PTGS1, PPARA, SLC01B1, ESR2, TYR, TAAR1, AAAS, CYP19A1, HSD11B1, SQLE, GLI2, GLI1, CYP3A5, CYP2D6, CES2, CA2, CA1, CA4, CA6, GPBAR1, FASN, MBOAT4, TOP2A, PTPN1, PRKCE, ESR2, AR, RORA, NR5A1, SHBG, ALDH5A1, ABAT, MET, AKR1B10, TOPBP1, CDC25A, DUSP3

**Table 3:** Respiratory disease associated Protein targets

Botanicals	Ligand	Protein targets
<i>Mimusops elengi</i> L. (72)	Quercetin	→EGFR, →KDR, →MET, →CDK6, →LCK, →IGF1R, →CDK1, →AURKB, →TTR, →TYR, →CDK5, →PI4KA, →ABCG2, →CA4, →CYP1A1, →PIK3CG, →AXL, →ALOX5, →NOX4, →MAOA, →ALOX15, →ADORA3, →AKR1B10, →CYP19A1
	Quercitol	→SRC, →CA4
	β-sitosterol- β-D-glucoside	→STAT3, →BCL2L1, →UBE2N, →F7, →TOPBP1, →AKR1B10, →POLL, →TOP2A, →IL2, →ABCC4
	Ursolic acid	→AKR1B10, →PTPRC, →ELANE, →ALOX15, →HSD11B2, →HSD11B1, →CDC25B, →PRKCH, →PPARA, →F7
	Stearic acid 3-(octadecyloxy) propyl ester	→PRKCA
	Hexadecanoic acid, methyl ester	→TLR2, →FFAR1, →DNM1, →FABP4
	10-Octadecenoic acid, methyl ester	→TOPBP1, →CACNA1S, →SCD, →FFAR4, →FFAR1, →FABP4
	Spinasterol	→CYP17A1, →CYP19A1, →PTPN2, →SREBF2, →AAAAS, →RORC
	β-carotene	→ADORA2B, →ADORA2A, →ADORA1, →ADORA3
	β-sitosterol	→CYP17A1, →CYP19A1, →PTPN2, →SREBF2, →RORC
<i>Kedrostis foetidissima</i> (Jacq.) Cogn. (78)	Quercetin-3-O-rhamnoside	→EGFR, →F10, →CHRM5, →TNF, →NOX4, →ADORA3, →RPS6KA3, →ADORA2A, →AKR1B10, →ALOX5, →CA4, →IL2
	7,10-Hexadecadienoicacid, methylester	→TOPBP1, →F7, →ATG4B, →FABP4, →SOAT1, →OXER1, →CYP19A1
	Eicosanoic acid, methyl ester(CAS)	→TLR2, →DNM1, →FFAR1, →AKR1B10, →FABP4
	Phytol acetate	→TRPV1 (VR1) →PRKCA, →NR3C1, →CYP17A1, →HSD11B1, →CYP19A1, →PRKCD,
	Curcubitacin	→AKR1C1, →ITGAL →ARHGAP35, →F10, →PLA2G2A, →PRSS1
	1H-Indole, 1-methyl-(CAS)	→HSP90AA1, →AAAAS, →HTR2C, →HTR2A, →HTR6, →DDC, →KIF23, →CA4
	Rutin	→IL2, →ADORA2A, →CA4, →ALOX5, →ADORA3, →AKR1B10, →CHRM5, →NOX4, →RPS6KA3, →F10, →TNF
	Docosanoic acid	→CDC25A, →FABP4, →AKR1B10, →TLR2, →DNM1, →FFAR1, →ALOX5
	9-Octadecenoic acid, methyl ester	→PPARA, →SCD, →FABP4, →FFAR4, →TOPBP1, →CYP19A1, →F7, →SOAT1, →ATG4B, →FFAR1
	1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester	→CTSK, →CTSB
<i>Artemisia vulgaris</i> L. (35)	Citrulline	→NOS3, →NOS1, →NOS2
	Chavicol	→TAARI
	Borneol	→RORA, →HSD11B1, →AKR1B10
	Osopulegol	→PRKCB, →PRKCA, →PRKCE, →TOP2A
	Myrtenol	→TOPBP1, →CDC25A, →AKR1B10, →TOP2A
	Camphor	→F12, →G6PD, →CYP19A1, →SOAT1, →TRPA1, →CA4, →FASN
	p-Cymene	→TAARI, →AAAAS
	1,8-Cineole	→HSD11B1, →CYP19A1
	α-Pinene	→PPARA, →CNR1, →CNR2
	β-Caryophyllene	→CNR1, →CNR2, →PPARA
(Z)-β-Farnesene	→MAOA	
Trans-2-hexenal	→TRPA1	
D-Limonene	→TRPA1, →PPARA, →CNR1, →CNR2	

[Red arrow (→) indicates the ligands inhibitory action. Green arrow (→) indicates the activator action of the ligand and Black arrow (→) indicates the unknown modulation]

**Search for Potential protein targets**

Protein targets for each phytoconstituent present in the herbal formulation MKA was identified from Binding database and presented in Table 2. In MKA, 110 protein targets were found for *Mimusops elengi* L., 91 targets for *Kedrostis foetidissima* (Jacq.) Cogn. and 50 protein targets for *Artemisia vulgaris* L. The combinatorial action of

these multiple ligands is the key factor enhancing the activity of Polyherbal formulation MKA.

**Disease associated target identification in Open target platform**

Respiratory disease associated protein targets of MKA were identified from Open target Platform and illustrated in Table 3. In MKA, 72/110

protein targets of *Mimusops elengi* L., 78/91 targets of *Kedrostis foetidissima* (Jacq.) Cogn. and 35/50 protein targets of *Artemisia vulgaris* L. were found to be the respiratory disease associated therapeutic targets. In Table 3, Red colored arrow (→) indicates the ligands inhibitory action, Green arrow (→) indicates the activator action of the ligand and Black arrow (→) indicates the unknown modulation].

**Ligand-target interactions in respiratory disease**

In MKA, β-sitosterol- β-D-glucoside has inhibitory action on Signal transducer and activator of transcription 3 (STAT3). STAT-3 inhibitors were found to reduce the production of cytokines responsible for lung inflammation [20]. EGFR signalling pathway inhibitors reduces the pathogenesis of Chronic lung disease [21]. Quercetin and Quercetin-3-O-rhamnoside acts as inhibitor of Epidermal Growth factor receptor (EGFR), which confirms its therapeutic potential in treating respiratory disease. There is dramatic increase in level of TRPV1 (VR1) and TRPA1 in airway cell in respiratory virus infection [22]. Phytol acetate has inhibitory action on TRPV1, which validates its therapeutic activity in treating respiratory virus infection. Camphor, Trans-2-hexenal and D-Limonene acts as inhibitor of TRPA1 validating its potential in acting against respiratory viral infection.

Carbonic anhydrase inhibitors functions as respiratory stimulant in patients with Chronic Obstructive Pulmonary disease (COPD) [23]. Quercetin, Quercitol, Quercetin-3-O-rhamnoside, 1H- Indole, 1-methyl- (CAS), rutin and camphor has inhibitory action on carbonic anhydrase. PPARA agonists has anti-inflammatory property and inhibits the formation of lung fibrosis [24]. Ursolic acid, 9-Octadecenoic acid, methyl ester, α-Pinene, β-Caryophyllene and D-

Limonene functions as activator of PPARA, which further proves the efficacy of MKA in alleviating the respiratory disease. Inhibition of KDR reduces the pathogenesis of lung cancer [25]. Quercetin has inhibitory action on KDR. The β-sitosterol-β-D-glucoside acts as inhibitor of STAT-3. Inhibition of SteroylCoA desaturase 1 (SCD1) induces apoptosis of lung cancer cells and reduce the pathogenesis of the disease [26]. 9-Octadecenoic acid, methyl ester and 10-Octadecenoic acid, methyl ester in MKA has inhibitory action on SCD1. The multi-targeted action of MKA is responsible for enhanced therapeutic property in fighting respiratory disease.

**Construction of Protein-protein network**

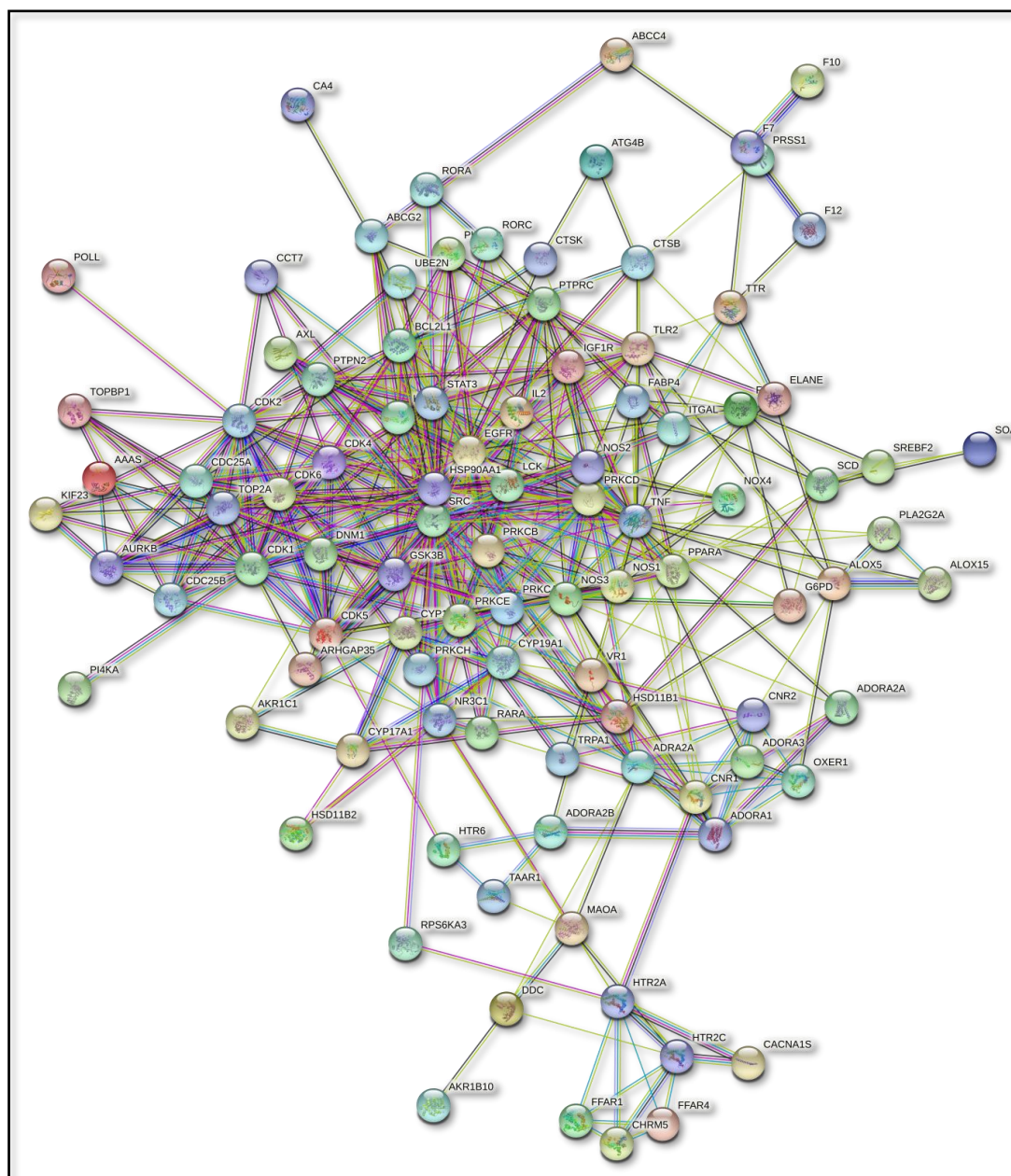
Multi-target interactions of MKA were illustrated in terms of PPI network in Figure 1.

The MKA network has 98 different protein targets interacting differently to produce therapeutic benefit in treating respiratory disease. In MKA network, 98 protein targets interact in different manner to produce the combined pharmacological effect. There are 460 metabolic interactions in MKA network. The KEGG pathway based functional enrichment analysis of MKA network was studied. The top 20 pathways involved in molecular mechanism of MKA is given in the Table 4.

This research proves the combinatorial action of multiple bioactive compounds present in herbal formulation MKA against respiratory disease. The molecular mechanism underlying the action of MKA herbal formulation was studied in terms of ligand-protein interaction and multi-target interactions.

**Table 4:** KEGG Pathway Functional Enrichment analysis

S. No	Pathway	Description	Count in Network	Strength	P-value
1	<a href="#">hsa00220</a>	Arginine biosynthesis	3 of 20	1.48	0.00096
2	<a href="#">hsa00360</a>	Phenylalanine metabolism	2 of 17	1.37	0.0095
3	<a href="#">hsa04750</a>	Inflammatory mediator regulation of TRP channels	10 of 92	1.34	1.00E-08
4	<a href="#">hsa05223</a>	Non-small cell lung cancer	6 of 66	1.26	1.74E-05
5	<a href="#">hsa04370</a>	VEGF signaling pathway	5 of 59	1.23	0.00012
6	<a href="#">hsa00330</a>	Arginine and proline metabolism	4 of 48	1.22	0.00071
7	<a href="#">hsa04540</a>	Gap junction	7 of 87	1.21	7.88E-06
8	<a href="#">hsa00380</a>	Tryptophan metabolism	3 of 40	1.18	0.004
9	<a href="#">hsa04066</a>	HIF-1 signaling pathway	7 of 98	1.15	1.28E-05
10	<a href="#">hsa04659</a>	Th17 cell differentiation	7 of 102	1.14	1.44E-05
11	<a href="#">hsa00591</a>	Linoleic acid metabolism	2 of 29	1.14	0.0205
12	<a href="#">hsa04020</a>	Calcium signaling pathway	12 of 179	1.13	1.51E-08
13	<a href="#">hsa04270</a>	Vascular smooth muscle contraction	8 of 119	1.13	4.95E-06
14	<a href="#">hsa04710</a>	Circadian rhythm	2 of 30	1.12	0.0215
15	<a href="#">hsa05222</a>	Small cell lung cancer	5 of 92	1.04	0.00069
16	<a href="#">hsa00590</a>	Arachidonic acid metabolism	3 of 61	0.99	0.0095
17	<a href="#">hsa04926</a>	Relaxin signaling pathway	6 of 130	0.96	0.00038
18	<a href="#">hsa04064</a>	NF-kappa B signaling pathway	4 of 93	0.93	0.0043
19	<a href="#">hsa03320</a>	PPAR signaling pathway	3 of 72	0.92	0.0132
20	<a href="#">hsa04620</a>	Toll-like receptor signaling pathway	3 of 102	0.77	0.0272



**Figure 1:** Protein-Protein Interaction network of MKA in respiratory diseases: The MKA network has average node degree of 9.39 and average local clustering coefficient of 0.579. There are 98 nodes and 460 edges. PPI enrichment p-value is less than 1.0e-16

## CONCLUSION

To conclude, bioactives in herbal formulation MKA, exert their therapeutic effects against respiratory disease by multiple mechanisms like STAT-3 inhibition by  $\beta$ -sitosterol- $\beta$ -D-glucoside, EGFR inhibition by Quercetin and Quercetin-3-O-rhamnoside, TRPA1 inhibition by D-Limonene, etc. The therapeutic benefit of MKA in treating respiratory disease is validated by its multi-targeted actions like inhibition of carbonic anhydrase by Quercetin, Quercitol, Quercetin-3-O-rhamnoside, 1H- Indole, 1- methyl- (CAS), rutin and camphor. It was evident that 98 protein targets interact differently in Protein-protein interaction network of MKA polyherbal formulation. The key target proteins were found to be EGFR, KDR, CA4, STAT-3, TRPA1, TRPV1 (VR1), PPARA and SCD1.

Thus, the network pharmacological approach was used to interpret the biological action of Polyherbal formulation MKA against respiratory diseases.

## Conflict of interest

Authors declare no conflict of interests.

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None.

## REFERENCES

1. Bateman ED, Bousquet J, FitzGerald M. Global strategy for asthma management and prevention: GINA executive summary. *European Respiratory Journal* 2008;31(1):143-78.
2. Ware LB, Matthay MA. The acute respiratory distress syndrome. *New England Journal of Medicine* 2000 May 4;342(18):1334-49.
3. Chung KF, Caramori G, Groneberg DA, Lamela J, Vega F, Blanco J, Hogg JC, Barnes PJ. Airway obstruction in chronic

- obstructive pulmonary disease [2](multiple letters). *New England Journal of Medicine* 2004 Sep 30;351(14):1459-61.
4. Drosten C, Günther S, Preiser W, Van Der Werf S, Brodt HR, Becker S, Rabenau H,
  5. Panning M, Kolesnikova L, Fouchier RA, Berger A. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *New England journal of medicine* 2003 May 15;348(20):1967-76.
  6. Ksiazek TG, Erdman D, Goldsmith CS, Zaki SR, Peret T, Emery S, Tong S, Urbani C, Comer JA, Lim W, Rollin PE. A novel coronavirus associated with severe acute respiratory syndrome. *New England journal of medicine* 2003 May 15;348(20):1953-66.
  7. Yuan H, Ma Q, Cui H, Liu G, Zhao X, Li W, Piao G. How can synergism of traditional medicines benefit from network pharmacology?. *Molecules* 2017 Jul;22(7):1135.
  8. Pool JL. Is it time to move to multidrug combinations?. *American journal of hypertension* 2003 Nov 1;16(S3):36S-40S.
  9. Chandran U, Mehendale NE, Tillu GI, Patwardhan BH. Network pharmacology: an emerging technique for natural product drug discovery and scientific research on ayurveda. *In Proc Indian Natn Sci Acad* 2015 Jun (Vol. 81, No. 3, pp. 561-8).
  10. Poongodi, T., Nazeema, T., & Ranjini, B. (2020). Enhanced Anti-Inflammatory Effect of Polyherbal Formulation (MKA) Comprising of Three Selected Plants in Lipopolysaccharide (LPS)-Induced Raw 264.7 Macrophage Cell Line. *Indian Journal of Pharmaceutical Sciences*, 82(4), 692-697.
  11. Poongodi T, & Nazeema T H. (2020). The Therapeutic effect of MKA on Bacterial Lipopolysaccharide (LPS) induced lipid peroxidation, cytosolic LDH leakage and mitochondrial membrane depolarization in RAW 264.7 Macrophages. *International Journal of Research in Pharmaceutical Sciences*, 11(4), 6250-6255.
  12. Azhagumurugan C, Rajan MK. GC-MS Analysis of Phytochemical Constituents and Nematicidal Activities of Leaf Extract of *Magilam, Mimusops elengi* L. *World Journal of Zoology* 2014;9(4):239-43.
  13. Rathinamala J, Anjana JC, Sruthy PB, Jayashree S. Phytochemical screening and GC MS analysis of bioactive compounds from *Mimusops elengi* L.. *Annals of Pharma Research* 2013;1(2).
  14. Kalaisezhiyen P, Sasikumar V. GC-MS evaluation of chemical constituents from methanolic leaf extract of *Kedrostis foetidissima* (Jacq.) Cogn. *Asian Journal of Pharmaceutical and Clinical Research* 2012;5(Suppl 4):77-81.
  15. Pavithra K, Saravanan G. A Review on Phytochemistry, Pharmacological Action, Ethanobotanical Uses and Nutritional Potential of *Kedrostis foetidissima* (Jacq.) Cogn. *Cardiovascular & Hematological Agents. Medicinal Chemistry*. 2020 May 1;18(1):5-20.
  16. Williams JD, Campbell MA, Jaskolka MC, Xie T. *Artemisia vulgaris* L. chemotypes. *Am J Plant Sci* 2013;4:1265-1269.
  17. Kim S, Thiessen PA, Bolton EE, Chen J, Fu G, Gindulyte A, Han L, He J, He S, Shoemaker BA, Wang J. PubChem substance and compound databases. *Nucleic acids research* 2016 Jan 4;44(D1):D1202-13.
  18. Gilson MK, Liu T, Baitaluk M, Nicola G, Hwang L, Chong J. BindingDB in 2015: a public database for medicinal chemistry, computational chemistry and systems pharmacology. *Nucleic acids research* 2016 Jan 4;44(D1):D1045-53.
  19. Koscielny G, An P, Carvalho-Silva D, Cham JA, Fumis L, Gasparyan R, Hasan S, Karamanis N, Maguire M, Papa E, Pierleoni A. Open Targets: a platform for therapeutic target identification and validation. *Nucleic acids research* 2017 Jan 4;45(D1):D985-94.
  20. Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-Cepas J, Simonovic M, Roth A, Santos A, Tsafou KP, Kuhn M. STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic acids research* 2015 Jan 28;43(D1):D447-52.
  21. Gavino AC, Nahmod K, Bharadwaj U, Makedonas G, Tweardy DJ. STAT3 inhibition prevents lung inflammation, remodeling, and accumulation of Th2 and Th17 cells in a murine asthma model. *Allergy* 2016 Dec;71(12):1684-92.
  22. Vallath S, Hynds RE, Succony L, Janes SM, Giangreco A. Targeting EGFR signalling in chronic lung disease: therapeutic challenges and opportunities. *Eur Respir J* 2014;44:513-522.
  23. Omar S, Clarke R, Abdullah H, Brady C, Corry J, Winter H, Touzelet O, Power UF, Lundy F, McGarvey LP, Cosby SL. Respiratory virus infection up-regulates TRPV1, TRPA1 and ASIC3 receptors on airway cells. *PLoS One* 2017;12(2).
  24. Adamson R, Swenson ER. Acetazolamide use in severe chronic obstructive pulmonary disease. pros and cons. *Annals of the American Thoracic Society* 2017 Jul;14(7):1086-93.
  25. Lakatos HF, Thatcher TH, Kottmann RM, Garcia TM, Phipps RP, Sime PJ. The role of PPARs in lung fibrosis. *PPAR research* 2007;2007.
  26. Mockus SM, Potter CS, Stafford GA, Ananda G, Hinerfeld D, Tsongalis GJ. Targeting KDR mutations in lung adenocarcinoma. *Cancer Res* 2015;75(15 Supplement);73-73.
  27. Hess D, Chisholm JW, Igal RA. Inhibition of stearylCoA desaturase activity blocks cell cycle progression and induces programmed cell death in lung cancer cells. *PloS one* 2010;5(6).

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